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Phenylpropanoids from the Immature Fruits of Black Nightshade (Solanum nigrum L.)

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Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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Original Research Article

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ABSTRACT

Black nightshade (*Solanum nigrum* L.) has been traditionally used as indigenous Chinese medicine. Phytochemical investigations of the immature fruits of *S. nigrum* have been carried out and nine phenylpropanoids (1-9) were obtained. Their structures were elucidated on the basis of spectroscopic and chemical methods, including MS, ¹H and ¹³C-NMR. They were identified as (7*S*, 8*R*)-4-[3-hydroxymethyl-5-(3- hydroxypropyl)-2,3-dihydrobenzofuran-2-yl]-2-methoxyphenol (1), (7*S*, 8*R*)- dihydrodehydroconifery alcohol (2), massonianoside A (3), butane-2,3-diol 2-O- (6-O-caffeoyl)- β -D-glucopyraniside (4), 4-[(6-O-(*E*)-caffeoyl)- β -D-glucopyranosyl] vanillic acid (5), (+)-isolariciresinol (6), *trans*-cinnamic acid (7), ferulic acid (8), and 4-hydroxy cinnamic acid (9). This study enriched the chemical constituents of black nightshade.

Keywords: Black nightshade; Solanum nigrum L.; immature fruits; chemical investigation; phenylpropanoids.

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1. INTRODUCTION

"Black nightshade (Solanum nigrum L.) belongs to the family of Solanaceae and is wildly distributed all over the world" [1]. "In China, it has been used as a common traditional Chinese medicine, which has the effects of clearing heat and detoxification, activating blood circulation and removing blood stasis, promoting water and swelling, and is mainly used to treat cold and fever. toothache, and cancers. Modern pharmacological studies indicated that S. nigrum exhibited a variety of biological activities including antiproliferative, anti-inflammatory [2,3], antiviral, hepatoprotective, and antioxidative activities" [4-6].

"S. nigrum contains steroidal saponins, steroidal alkaloid glycosides and phenolic compounds in the previous chemical studies" [7-10]. *"Steroidal alkaloid glycosides are the main chemical components of Solanum species, which possess various pharmacological activities such as antiproliferative and anti-inflammatory properties" [11,12]. Currently, most of the chemical investigations on <i>S. nigrum* were focused on the aerial parts, while the bioactive components of its unripe berries are still unclear.

In this study, the phytochemicals from the immature fruits of *S. nigrum* has been carried out and nine phenylpropanoids were obtained.

2. MATERIALS AND METHODS

2.1 General Experimental Procedures

MS spectra were obtained on an Acquity UPLC-Q-TOF Microsystem (Waters Co., Milford, MA). NMR spectra were taken on a Bruker Avance III 500 MHz spectrometer (Bruker, Switzerland). ODS packed column (40–60 µm, Merck KGaA, Darastadt, Germany) and column chromatography was employed on silica gel (Anhui Liangchen Silicon Source Material Co. Ltd, Lu'an, China). All other analytical chemicals and reagents were purchased from Sinopharm Chemical Reagent Co. Ltd. (Shanghai, China).

2.2 Plant Materials

The immature fruits of *S. nigru*m were purchased from Haerbin (Heilongjiang province, China) and dried at room temperature in the shade. The voucher specimen has been deposited in the School of Pharmacy, Guangdong Pharmaceutical University, China.

2.3 Extraction and Isolation

Dried immature fruits of *S. nigru*m (2.5 kg) were extracted with 70% EtOH (v/v, 15 L × 3) for 2 h. The ethanol-free suspension was subjected to a D101 macroporus resin column (80 × 1100 mm), and eluted with H₂O, 10% MeOH, 30% MeOH, 50% MeOH, 70% MeOH and MeOH to give six fractions (I-VI). The 70% MeOH elution (fraction V, 45.1 g) was separated by a silica gel chromatography column (200-300 mesh, 1100 g) into 15 fractions (V-1 to V-15) with a CHCl₃-MeOH gradient (100:1 to 0:1, v/v).

Compounds 7 (16.2 mg), 8 (10.1 mg), and 9 (16.0 mg) were obtained from fraction V-8 followed by an ODS MPLC, gradiently eluted with MeOH-H₂O (1:9 to 10:0, v/v). Fraction V-6 was applied to an ODS MPLC eluted with a gradient of MeOH-H₂O (1:9 to 10:0, v/v) to afford eight subfractions (V-6-1 to V-6-8). Subfraction V-6-5 was further purified by a semi-preparative HPLC to obtained compounds 1 (8.2 mg) and 3 (7.3 mg). Fraction V-7 was subjected to an ODS MPLC and eluted with MeOH-H₂O gradient (1:9 to 10:0, v/v) to afford ten fractions (V-7-1 to IV-7-10). Subfraction V-7-6 was further separated by an ODS MPLC and semi-preparative HPLC to obtained compounds 2 (10.7 mg), 4 (6.3 mg), 5 (9.0 mg), and 6 (5.6 mg), respectively.

3. RESULTS AND DISCUSSION

The seventy percentage ethanol extract of the immature fruits of *S. nigrum* was separated successively by column chromatography on D101 macroporus resin, silica gel, ODS MPLC, and preparative HPLC, to afford nine phenylpropanoids (**1-9**) (Fig. 1). Their structures were elucidated on the basis of spectroscopic data, including MS, and ¹H and ¹³C-NMR.

Compound 1, brownish oil, was a blue fluorescence under 365 nm after TLC development. Its molecular formula was determined as $C_{19}H_{22}O_5$ based on its ESI-MS with the ion *m*/*z* 353 [M+Na]⁺, ¹H and ¹³C-NMR.

In the ¹H-NMR, protons at $\delta_{\rm H}$ 7.01 (1H, d, *J*=1.8 Hz, H-2), 6.87 (1H, dd, *J*=8.2, 1.8 Hz, H-6) and 6.80 (1H, d, *J*=8.2 Hz, H-5) consisted an ABX coupling system of in a 1,3,4-trisubstituted benzene ring. Signals at $\delta_{\rm H}$ 7.13 (1H, s, H-2'), 7.00 (1H, d, J=8.1 Hz, H-6') and 6.70 (1H, d, J=8.1 Hz, H-6') and 6.70 (1H, d, J=8.1 Hz, H-5') came from another ABX coupling system benzene ring in the molecule. $\delta_{\rm H}$ 5.48 (1H, d, *J*=6.4 Hz, H-7) was a proton linked to oxygenated carbon. $\delta_{\rm H}$ 3.80 (3H, s, 3-OCH₃) was a methoxy group.

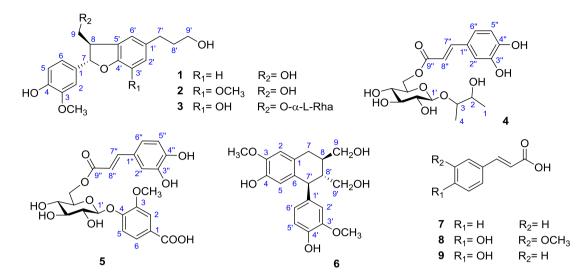


Fig. 1. Structures of the phenylpropanoids 1-9

¹³C-NMR gave 19 carbon signals (Table 1). In aromatic region ($δ_{\rm C}$ 159.1 to 109.5), there were 12 carbon signals, which were two benzene ring units. Carbon signal at $δ_{\rm C}$ 56.4 in high field region is a methoxy carbon signal, which was consistent with the information given by ¹H-NMR. $δ_{\rm C}$ 87.8, 54.8 and 64.8 were deduced as the carbon signals on the furan ring, and $\delta_{\rm C}$ 36.1, 32.4 and 61.9 were form a hydroxypropyl group. Taken together, compound **1** was speculated a benzodihydrofuran lignin. Based on above analysis and the literature [13], compound **1** was identified as (7*S*, 8*R*)-4-[3-hydroxymethyl-5-(3hydroxypropyl)-2,3-dihydrobenzofuran-2-yl]-2methoxyphenol.

The molecular formula of compound **2** was determined as $C_{20}H_{24}O_6$ based on its ESI-MS with the ion m/z 383 [M+Na]⁺, ¹H and ¹³C-NMR. In the ¹H-NMR, protons at δ_H 7.03 (1H, d, *J*=1.8 Hz, H-2), 6.88 (1H, dd, *J*=8.1, 1.8 Hz, H-6) and 6.81 (1H, d, *J*=8.1 Hz, H-5) consisted an ABX coupling system of in a 1,3,4-trisubstituted benzene ring. Signals at δ_H 6.74 (1H, s, H-2') and 6.76 (1H, d, *J*=8.1 Hz, H-6') came from another benzene ring in the molecule. δ_H 5.50 (1H, d, *J*=6.4 Hz, H-7) was a proton linked to oxygenated carbon. δ_H 3.82 (3H, s, 3-OCH₃) and 3.81 (3H, s, 3'-OCH₃) were two methoxy groups.

In the ¹³C-NMR (Table 1), there were 20 carbon signals, including 12 carbon signals in aromatic region ($\delta_{\rm C}$ 148.4 to 110.6), which were two benzene ring units. Carbon signals at $\delta_{\rm C}$ 56.4 and 56.3 in high field region were two methoxy carbons, which were consistent with the information given by ¹H-NMR. $\delta_{\rm C}$ 88.3, 55.2 and 64.8 were deduced as the carbon signals on the furan ring, and δ_c 36.1, 32.8 and 61.8 were form a hydroxypropyl group. Therefore, compound **2** was speculated a benzodihydrofuran lignin. Based on above analysis and the literature [14], compound **2** was identified as (7*S*, 8*R*)dihydrodehydroconifery alcohol.

Compound 3, brownish oil, was а blue fluorescence under 365 nm after TLC development. lts molecular formula was determined as C₂₅H₃₂O₁₀ based on its ESI-MS with the ion m/z 515 [M+Na]⁺, ¹H and ¹³C-NMR.

In the ¹H-NMR, protons at $\delta_{\rm H}$ 6.93 (1H, d, *J*=1.3 Hz, H-2), 6.86 (1H, dd, *J*=8.2, 1.3 Hz, H-6) and 6.81 (1H, d, *J*=8.2 Hz, H-5) consisted an ABX coupling system of in a 1,3,4-trisubstituted benzene ring. Signals at $\delta_{\rm H}$ 6.50 (2H, br.s, H-2', 6') came from another benzene ring in the molecule. $\delta_{\rm H}$ 5.37 (1H, d, *J*=6.6 Hz, H-7) was a proton linked to oxygenated carbon. $\delta_{\rm H}$ 3.76 (3H, s, 3-OCH₃) and 1.13 (3H, d, *J*=6.2 Hz, H-6") were a methoxy and methyl groups in the higher field. $\delta_{\rm H}$ 4.62 (1H, d, *J*=0.7 Hz, H-1") was deduced as a terminal proton signal of sugar.

In the ¹³C-NMR (Table 1), there were 25 carbon signals, including 12 carbon signals in aromatic region ($\delta_{\rm C}$ 144.7 to 110.2), which were two benzene ring units. Carbon signal at $\delta_{\rm C}$ 55.5 in high field region was a methoxy carbon, which was consistent with the information given by ¹H-NMR. Compared with the NMR data of compound **2**, it is speculated that compound **3** is a benzodihydrofuran lignin with rhamnose substitution at C-9 position. Based on above analysis and the literature [15], compound **3** was identified as massonianoside A.

NO.	1 ^a	2 ^a	3 ^a	4 ^b	5 ^b	6 ^ª
1	134.9	134.8	132.6	15.1	123.7	128.5
2 3	110.5	110.6	110.2	78.6	112.7	111.8
3	148.5	147.4	146.4	68.9	149.1	146.5
4	147.2	148.4	147.7	19.5	151.6	145.3
5 6	115.6	115.7	114.8		114.9	116.9
6	119.6	119.6	118.6		123.4	138.5
7	87.8	88.3	87.2		167.9	33.8
8	54.8	55.2	50.9			40.4
9 1'	64.8	64.8	68.9			65.9
1'	135.4	130.1	128.3	101.3	100.5	133.9
2'	125.8	113.9	115.4	73.4	73.2	113.6
3'	129.1	147.2	140.8	76.6	75.8	148.3
4'	159.1	144.9	144.7	70.5	70.3	145.9
5'	109.5	136.4	135.2	73.8	73.7	115.6
6'	129.3	117.7	116.2	63.8	64.0	122.8
7'	32.4	36.1	34.8			48.1
8'	36.1	32.8	31.5			48.4
9'	61.9	61.8	60.3			62.1
1"			100.2	125.6	125.9	
2"			70.9	113.9	115.3	
3"			72.0	145.8	145.3	
4"			70.6	148.7	147.4	
5"			68.6	115.9	116.3	
6"			18.1	121.5	120.5	
7"				145.4	144.1	
8"				114.9	115.2	
9"				166.5	165.6	
3-OCH₃	56.4	56.4	55.5		55.6	56.2
3'-OCH ₃		56.3	<u>k</u>	43	NMR in DMSO-0	56.3

Table 1. ¹³C NMR data of compounds 1-6 (δ in ppm and J in Hz)

^a 151 MHz for ¹³C NMR in Acetone-d₆.; ^b 101 MHz for ¹³C NMR in DMSO-d₆

Compound **4** was brownish oil and exhibited a blue fluorescence under 365 nm after TLC development. Its molecular formula was determined as $C_{19}H_{26}O_{10}$ based on its ESI-MS with the ion m/z 437 [M+Na]⁺, ¹H and ¹³C-NMR.

In the ¹H-NMR spectrum, signals at $\delta_{\rm H}$ 7.50 (1H, d, J=15.6 Hz, H-7") and 6.26 (1H, d, J=15.6 Hz, H-8") were speculated to be the proton signals on the trans carbon-carbon double bond. The aromatic region had three proton signals at $\delta_{\rm H}$ 7.04 (1H, s, H-2"), 6.99 (1H, d, J=7.8 Hz, H-5") and 6.76 (1H, d, J=7.4 Hz, H-6"). Signal at $\delta_{\rm H}$ 4.26 (1H, d, J=7.7 Hz, H-1') was the terminal proton signal of sugar. $\delta_{\rm H}$ 4.40~2.98 has 8 proton signals, which were proton signals on oxymethylene or oxymethylene. $\delta_{\rm H}$ 1.07 (3H, d, J=6.0 Hz, H-1) and 0.99 (3H, d, J=6.0 Hz, H-4) were two methyls in the high field.

¹³C-NMR spectrum indicated 19 carbon signals, and $\delta_{\rm C}$ 166.5 was an ester carbonyl in the lower field (Table 1). Signals at $\delta_{\rm C}$ 166.5, 148.7, 145.8,

145.4, 125.6, 121.5, 115.9, 114.9 and 113. 9 consisted of a caffeic acyl group. Meanwhile, compound **4** has a glucose unit, which the carbon signals at $\delta_{\rm C}$ 101.3, 73.4, 76.6, 70.5, 73.8 and 63.8. The remaining four carbon signals at $\delta_{\rm C}$ 78.6, 68.9, 19.5 and 15.1 formed a 3-hydroxy-2-butoxy group. Based on above analysis and the literature [16], compound **4** was identified as butane-2, 3-diol 2-O-(6-O-caffeoyl)- β -D-glucopyraniside.

Compound **5** was brownish oil and exhibited a blue fluorescence under 365 nm after TLC development. Its molecular formula was determined as $C_{23}H_{24}O_{12}$ based on its ESI-MS with the ion m/z 491 [M–H]⁻, ¹H and ¹³C-NMR.

In the ¹H-NMR spectrum, signals at $\delta_{\rm H}$ 7.48 (1H, d, J=15.7 Hz, H-7") and 6.36 (1H, d, J=15.7 Hz, H-8") were speculated to be the proton signals on the *trans* carbon-carbon double bond. The aromatic region had six proton signals at $\delta_{\rm H}$ 7.40 (1H, dd, *J*=8.2, 1.7 Hz, H-6), 7.38 (1H, d, *J*=1.7

Hz, H-2), 7.37 (1H, d, *J*=1.8 Hz, H-2"), 7.18 (1H, dd, *J*=8.2, 1.8 Hz, H-6"), 6.84 (1H, d, *J*=8.2 Hz, H-5) and 6.77 (1H, d, *J*=8.2 Hz, H-5"), which speculated that the compound had two 1,3,4-trisubstituted benzene rings. Signal at $\delta_{\rm H}$ 5.08 (1H, d, *J*=7.3 Hz, H-1') was the terminal proton signal of sugar. $\delta_{\rm H}$ 3.73 (3H, s, 3-OCH₃) was a methyoxyl group in the high field.

The ¹³C-NMR spectrum gives 23 carbon signals (Table 1). Compared with the NMR data of compound **4**, compound **5** contained a caffeic acyl segment and a glucose unit. The chemical shifts of C-1' and C-6' of the glucose shifted to the low field, which speculated that there were substituted on C-1' and C-6'. Meanwhile, compound **5** had a vanillic acid fragment with substitutions at position 4. Based on above analysis and the literature [17], compound **5** was identified as $4-[(6-O-(E)-caffeoyl)-\beta-D-glucopyranosyl]$ vanillic acid.

Compound **6** was brownish oil and its molecular formula was determined as $C_{20}H_{24}O_6$ based on its ESI-MS with the ion *m/z* 359 [M–H]⁻, ¹H and ¹³C-NMR. In the ¹H-NMR spectrum, signals in aromatic region at δ_H 6.78 (1H, s, H-5 '), 6.66 (1H, s, H-2) and 6.22 (1H, s, H-5) were protons on the benzene ring. In the high field, δ_H 3.78 (3H, s, 3-OCH3) and 3.81 (3H, s, 3'-OCH3) were two methoxy signals.

The ¹³C-NMR spectrum gave 20 carbon signals (Table 1), and there had 12 carbon signals in the aromatic region, which were two benzene ring units. $\delta_{\rm C}$ 56.2 and 56.3 were two methoxy groups, which were consistent with the information given by ¹H-NMR spectrum. The remaining six carbon signals at $\delta_{\rm C}$ 65.9, 62.1, 48.4, 48.1, 40.4 and 33.78 were speculated that it was the alkyl carbon signal on the C₃ skeleton of phenylpropanoid. Based on above analysis and the literature [18], compound **6** was identified as (+)-isolariciresinol.

Compounds **7** to **9** were identified as *trans*cinnamic acid (**7**) [19], ferulic acid (**8**) [20], and 4hydroxy cinnamic acid (**9**) [21], respectively, based on their spectroscopic analysis.

4. CONCLUSIONS

Further chemical investigation of the immature fruits of *S. nigrum* led to the isolation of nine phenylpropanoids. Their structures were elucidated on the basis of spectroscopic and chemical methods. They were identifies as

compound was identified as (7S, 8R)-4-[3hydroxymethyl-5-(3-hydroxypropyl) -2,3dihydrobenzofuran-2-yl]-2-methoxyphenol, (7S, 8R)-dihydrodehydroconifery alcohol, massonianoside A (**3**), butane-2,3-diol 2-O-(6-Ocaffeoyl)- β -D- glucopyraniside, 4-[(6-O-(*E*)caffeoyl) - β -D-glucopyranosyl] vanillic acid, (+)isolariciresinol, *trans*-cinnamic acid, ferulic acid, and 4-hydroxy cinnamic acid.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. Wang FZ, Tang J. Flora of China. Beijing, China: Science Publishing House. 1980;76-9.
- Ding X, Zhu FS, Li M, Gao SG. Induction of apoptosis in human hepatoma SMMC-7721 cells by solamargine from *Solanum nigrum* L. J Ethnopharmacol. 2012; 139(2):599-604.
 DOI: 10.1016/j.jep.2011.11.058, PMID 22172325.
- Zakaria ZA, Gopalan HK, Zainal H, Mohd Pojan NHM, Morsid NA, Aris A, et al. Antinociceptive, anti-inflammatory and antipyretic effects of *Solanum nigrum* chloroform extract in animal models. Yakugaku Zasshi. 2006;126(11):1171-8. DOI: 10.1248/yakushi.126.1171, PMID 17077618.
- Javed T, Ashfaq UA, Riaz S, Rehman S, Riazuddin S. In-vitro antiviral activity of *Solanum nigrum* against hepatitis C virus. Virol J. 2011;8:26.
 DOI: 10.1198(4742.422X.8.26.

DOI: 10.1186/1743-422X-8-26, PMID 21247464.

5. Hsieh CC, Fang HL, Lina WC. Inhibitory effect of *Solanum nigrum* on

thioacetamide-induced liver fibrosis in mice. J Ethnopharmacol. 2008;119(1): 117-21.

DOI: 10.1016/j.jep.2008.06.002, PMID 18606216.

- Aly YS, Shallan MA. Antioxidant properties of wild Solanum nigrum ripe fruit. Planta Med. 2011;77(12):1367. DOI: 10.1055/s-0031-1282627
- Zhou XL, He XJ, Wang GH, Gao H, Zhou GX, Ye WC, et al. Steroidal saponins from Solanum nigrum. J Nat Prod. 2006;69(8):1158-63.
 DOI: 10.1021/np060091z, PMID 16933867.
- Wang Y, Xiang L, Yi X, He X. Potential anti-inflammatory steroidal saponins from the berries of *Solanum nigrum* L. (European Black nightshade). J Agric Food Chem. 2017;65(21):4262-72. DOI: 10.1021/acs.jafc.7b00985, PMID 28486801.
- Xiang L, Wang Y, Yi X, He X. Antiinflammatory steroidal glycosides from the berries of *Solanum nigrum* L. (European black nightshade). Phytochemistry. 2018;148:87-96. DOI: 10.1016/j.phytochem.2018.01.019,

PMID 29421515.

- Xiang L, Wang Y, Yi X, He X. Steroidal alkaloid glycosides and phenolics from the immature fruits of *Solanum nigrum*. Fitoterapia. 2019;137:104268.
 DOI: 10.1016/j.fitote.2019.104268, PMID 31306720.
- Wang Y, Xu J, Wang Y, Xiang L, He X. S-20, a steroidal saponin from the berries of black nightshade, exerts anti-multidrug resistance activity in K562/ADR cells through autophagic cell death and ERK activation. Food Funct. 2022;13(4): 2200-15. DOI: 10.1039/d1fo03191k, PMID

35119449.

12. Emmanuel S, Ignacimuthu S, Perumalsamy R, Amalraj T. Antiinflammatory activity of *Solanum trilobatum.* Fitoterapia. 2006;77(7-8):611-2. DOI: 10.1016/j.fitote.2006.09.009, PMID 17056205.

 Sinkkonen J, Karonen M, Liimatainen J, Pihlaja K. Lignans from the bark extract of *Pinus sylvestris* L. Magn Reson Chem. 2006;44(6):633-6. DOI: 10.1002/mrc.1780, PMID 16489551.

Hanawa F, Shiro M, Hayashi Y. Heartwood constituents of *Betula maximowicziana*. Phytochemistry. 1997;45(3):589-95. DOI: 10.1016/S0031-9422(96)00885-0

- Bi YF, Zheng XK, Feng WS, Zhang YZ, Ji CR. Isolation and structure identification of ligan glycosides from pine needles of Pinus massonniana Lamb. Yao Xue Xue Bao. 2002;37(8):626-9. PMID 12567777.
- Yang PF, Yang YN, Feng ZM, Jiang JS, Zhang PC. Six new compounds from the flowers of *Chrysanthemum morifolium* and their biological activities. Bioorg Chem. 2019;82:139-44.
 DOI: 10.1016/j.bioorg.2018.10.007, PMID 30321776.
- Wang H, Zhao W, Choomuenwai V, Andrews KT, Quinn RJ, Feng Y. Chemical investigation of an antimalarial Chinese medicinal herb *Picrorhiza scrophulariiflora*. Bioorg Med Chem Lett. 2013;23(21): 5915-8. DOI: 10.1016/j.bmcl.2013.08.077, PMID 24035096.
- Okuyama E, Suzumura K, Yamazaki M. Pharmacologically active components of todopon puok (*Fagraea racemosa*), a medicinal plant from Borneo. Chem Pharm Bull (Tokyo). 1995;43(12):2200-4. DOI: 10.1248/cpb.43.2200, PMID 8582023.
- 19. Feng M, Wang S, Zhang X. Chemical constituents in fruits of *Lycium barbarum*. Chin Trad Herb Drugs. 2013;44:265-8.
- Yang X, Xu L, Yang S. Organic acid constituents from the stem of *Securidaca inappendiculata* Hassk. China J. Chin. mat. Med. 2001;26:258-60.
- 21. Pyo M, Koo Y, Yun-Choi H. Anti-platelet effect of the phenolic constituents isolated from the leaves of *Magnolia obovata*. Nat Prod Sci. 2002;8:147-51.

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