

Efficacy of aqueous extracts of some solanaceous plants on juveniles mortality and egg hatching on the root knot nematode, *Meloidogyne javanica* under laboratory conditions

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Abstract

Aqueous extracts of black nightshade, *Solanum nigrum*; deadly nightshade, *Atropa belladonna*; Egyptian henbane, *Hyoscyamus muticus*; thorn apple, *Datura innoxia*; ashwagndha, *Withania somnifera* leaves and fruits of chili pepper, *Capsicum frutescens* were assessed on juvenile mortality and egg hatching of the root knot nematode, *Meloidogyne javanica* *in vitro*. The nematode mortalities were in the range of (54.33 to 100.0 %) in concentrations S/16, S/8 and S/4 compared to distilled water (1.33 %) and increased with the increase of the exposure time from 24 to 72 hrs. Deadly nightshade and chili pepper extracts were the most effective against second stage juveniles of nematode at concentration of S/4, followed by aqueous extracts of ashwagndha, thorn apple, black nightshade. Result showed that, the highest reduction in hatching inhibition percentage was recorded with concentration (S/4) of ashwagndha (97.63 %) followed by Egyptian henbane, deadly nightshade, black nightshade, thorn apple and chili pepper with inhibition percentages as (95.42, 94.60, 92.87, 79.61 and 67.97%) respectively.

Keywords: solanaceous, *Meloidogyne javanica*, aqueous extracts, hatching and mortality.

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1. Introduction

Plant parasitic nematodes are one of the major problems in many parts of the world as well as soils of Nile valley and newly reclaimed sandy soils in desert of Egypt (Abd-Elgawad, 2013; Abd-Elgawad *et al.*, 2012; Oteifa and El-Ginidi, 1982). Nowadays, nematologists all over the world are keeping searching for alternative control measures to avoid soil pollution with chemical nematicides and subsequently the hazardous effects due to its residues. Resistant cultivars, plant extracts and manuring the soil with natural organic materials are commended as alternative methods of agricultural pest control in integrated pest management programs. The nematicidal activity of botanical extracts of chili pepper fruits (*Capsicum frutescens*) were evaluated on the root-knot nematode, *M. javanica* in tomato plants in the greenhouse, and in a second step, to compare the best extracts for the reduction of the number of eggs and root galls with two products containing capsaicin, capsaicoids and allyl isothiocyanate (Neves *et al.*, 2009). the effect of leaf extract of (*Withania somnifera*) and (*Datura metel*) were evaluated by observing number of knots, shoot length, fresh shoot weight, root length and fresh root weight of the plant (Singh and Devi, 2012). The effects of plant extracts from *Capsicum frutescens*, *Hyoscyamus niger* were evaluated against RKNs on eggs, egg masses and J₂s of the *M. incognita* in laboratory conditions (Kepenekci *et al.*, 2016). Leaf aqueous extracts of *Datura metel*, *Datura innoxia* were studied against the most devastating root-knot nematode *M. incognita*

(Nandakumar *et al.*, 2017). *Solanum nigrum* (seeds) and *Datura stramonium* (shoots) were investigated against root-knot nematodes in terms of J₂ paralysis and egg hatch inhibition (Oplos, 2018). The objective of this study was to test nematicidal activity of aqueous extracts representing six solanaceous plants for their potential toxicity against eggs and juveniles of the root knot nematode, *M. javanica* under laboratory conditions.

2. Materials and methods

Leaves of five solanaceous plants: black nightshade, *Solanum nigrum*; deadly nightshade, *Atropa belladonna*; Egyptian henbane, *Hyoscyamus muticus*; thorn apple, *Datura innoxia*; ashwagandha, *Withania somnifera* leaves and fruits of chili pepper, *Capsicum frutescens* were used. Plants materials collected randomly from plants grown in the experimental farm and gardens of Al-Azhar University, Assiut, Egypt except Egyptian henbane was collected from New Valley governorate. Plants parts were washed thoroughly under running tap water, cut into small pieces, shade dried and used for extraction. Dried plant materials homogenized to a fine powder then stored in airtight bottles. Fifty grams of each plant powder were extracted with 500 ml of distilled water for 24 hrs. The suspension was filtered with Whatman No. 1 filter paper for each plant extract and considered as a standard solution (S) and diluted to (S/4), (S/8) and (S/16). The culture of root knot nematode,

Meloidogyne javanica was maintained on potted eggplant planted inside the greenhouse. The infected plants uprooted, carefully washed in running tap water and egg-masses collected into Petri dishes containing distilled water. The evaluation was carried out in 5 cm clean Petri dishes in 3 replications for each treatment. The Petri dishes with distilled water was taken as control. 100-second stage juveniles (J₂s) of *M. javanica* were suspended in 10 ml of different extracts with dilutions S/16, S/8 and S /4. All Petri dishes were kept at ambient temperature at 28 ± 5 °C. After 24, 48 and 72 hrs, the dead J₂s were counted and the percentage of juveniles' mortality were calculated according to the following formula:

$$\text{Juveniles mortality \%} = \frac{\text{No. juveniles dead in the treatment}}{\text{Total No. juveniles were used in the treatment}} \times 100$$

Five egg-masses medium size handpicked from the galls of eggplant were placed in each of Petri dishes containing 5 ml of extracts with dilutions of S/8 and S /4. Egg-masses kept in distilled water served as control. Each treatment was replicated 3 times. After 7 days exposure, the numbers of juveniles hatched were counted and the percentages of egg hatched, and Inhibition were calculated using the following formulas:

$$\text{Eggs hatched \%} = \frac{\text{No. Juveniles hatched in the treatment}}{\text{No. juveniles hatched in control}} \times 100$$

$$\text{Inhibition of egg hatching \%} = \frac{\text{No. juveniles hatched in control} - \text{No. juveniles hatched in treatment}}{\text{Number of juveniles hatched in control}} \times 100$$

The randomized complete design was used in the experimental. Data were subjected to analysis of variance (ANOVA) using SPSS statistics. The

means were compared according to Duncan's multiple range tests at $P \leq 0.05$ (Duncan, 1955).

3. Results

The aqueous extracts of black nightshade (*Solanum nigrum*), deadly nightshade (*Atropa belladonna*), Egyptian henbane (*Hyoscyamus muticus*), chili pepper (*Capsicum frutescens*), thorn apple (*Datura innoxia*) and ashwagndha (*Withania somnifera*) were highly nematicidal effect to second stage juveniles of *M. javanica* *in vitro* (Table 1). The nematode mortality was in the range of (54.33 to 100.0 %) in concentrations S/16, S/8 and S /4 % compared to distilled water (1.33 %). Results revealed that, the nematode mortality with plant extracts were increased with the increase of the exposure time from 24 to 72 hrs. Deadly nightshade and chili pepper extracts were more effective against J₂ of nematode at concentration S/4 followed by aqueous extracts of ashwagndha, thorn apple, black nightshade and Egyptian henbane were (100, 100, 99, 98.33, 93 and 75.67 %), respectively, of exposure time 72 hrs. On the other hand, concentration S/16 recorded the lowest rate of juveniles mortalities in Egyptian henbane (54.33%), black nightshade (65.33%), ashwagndha (88%), chili pepper (89%), thorn apple (94.33%) and deadly nightshade (97.33%) respectively. Data presented in Table (2) indicated that, concentration (S/4) of aqueous extracts was the highest of percentage hatch

inhibition in ashwagndha (97.63 %) followed by Egyptian henbane, deadly nightshade, black nightshade, thorn apple and chili pepper with percentage inhibition of egg hatching (95.42, 94.60, 92.87, 79.61 and 67.97%) respectively. While, aqueous extracts of plants with

concentration S/8 showed the highest inhibition of egg-masses hatching in deadly nightshade (89.99 %) followed by ashwagndha (88.21%), Egyptian henbane (84.42%), black nightshade (78.64%), thorn apple (72.23%) and chili pepper (67.97%) respectively.

Table (1): Effect of aqueous extracts of six solanaceous plants on percentage of second stage juveniles mortality of the root-knot nematode, *Meloidogyne javanica*.

Materials	Conc.	Juveniles mortality after hrs (%)		
		24	48	72
Black nightshade (<i>Solanum nigrum</i>)	S/16	37.33 d	46.67 de	65.33 cd
	S/8	41.67 cd	63.33 bc	75.00 bc
	S/4	67.33 bc	75.67 b	93.00 a
Deadly nightshade (<i>Atropa belladonna</i>)	S/16	64.67 bc	26.00 f	97.33 a
	S/8	77.33 ab	30.00 f	98.33 a
	S/4	94.67 a	34.00 ef	100.00 a
Egyptian henbane (<i>Hyoscyamus muticus</i>)	S/16	54.67 bcd	45.33 de	54.33 d
	S/8	60.33 bcd	66.00 bc	66.67 cd
	S/4	76.67 ab	74.67 b	75.67 bc
Chili pepper (<i>Capsicum frutescens</i>)	S/16	46.00 cd	53.33 cd	89.00 ab
	S/8	97.33 a	98.00 a	99.00 a
	S/4	99.00 a	100.00 a	100.00 a
Thorn apple (<i>Datura innoxia</i>)	S/16	54.00 bcd	47.33 de	94.33 a
	S/8	59.00 bcd	93.67 a	96.00 a
	S/4	79.33 ab	98.33 a	98.33 a
Ashwagndha (<i>Withania somnifera</i>)	S/16	59.67 bcd	77.33 b	88.00 ab
	S/8	76.00 ab	91.67 a	93.33 a
	S/4	78.00 ab	95.33 a	99.00 a
Distilled Water (Control)		0.67 e	1.00 g	1.33 e

Values in each column followed by the same letters are no significant at P<0.05 according to Duncan's multiple-range test.

Table (2): Effect of aqueous extracts of six solanaceous plants on hatching of root-knot nematode, *Meloidogyne javanica* egg masses.

ds	Dose	Mean	Juveniles hatched (%)	Inhibition (%)
Black nightshade (<i>Solanum nigrum</i>)	S/8	345 cde	21.36	78.64
	S/4	115 fg	7.13	92.87
Deadly nightshade (<i>Atropa belladonna</i>)	S/8	128 fg	7.94	92.06
	S/4	87 fg	5.4	94.60
Egyptian henbane (<i>Hyoscyamus muticus</i>)	S/8	252 ef	15.58	84.42
	S/4	74 g	4.58	95.42
Chili pepper (<i>Capsicum frutescens</i>)	S/8	498 bc	30.78	69.22
	S/4	551 b	32.03	67.97
Thorn apple (<i>Datura innoxia</i>)	S/8	449 bcd	27.77	72.23
	S/4	329 de	20.39	79.61
Ashwagndha (<i>Withania somnifera</i>)	S/8	190 efg	11.79	88.21
	S/4	38 g	2.37	97.63
Distilled Water (Control)		1617 a	100	0.00

Values in each column followed by the same letters are no significant at P<0.05 according to Duncan's multiple-range test.

4. Discussion

In conclusion, the assessment results of land capability and land suitability for the selected crops helps in planning sustainable agriculture programs. Integration between geographic information systems (GIS), ASLE, and MicroLIES programs was undertaken in these soils to assess the land performance. The results of these soils indicated that the major soil limitations are soil texture, soil salinity, and low soil fertility characteristics, which can be improved using good management practices such as adding organic matter, fertilizers for upgrade the fertility, leaching the excess salt, and good agriculture practices for crops. These improvements will develop the potential suitability. Ultimately, from this study, it can be mentioned that the geostatistical approach and GIS are effective and strong tools for land capability and suitability studies and hence for sustainable planning of land use.

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