

International Journal of Plant & Soil Science

34(23): 1166-1173, 2022; Article no.IJPSS.93410 ISSN: 2320-7035

### Physiological Traits as the Primary Tool for Screening Salt Tolerance in Rice

K. Vanitha <sup>a¥\*</sup>, P. Shanthi <sup>b#</sup> and K. Subrahmaniyan <sup>c†</sup>

<sup>a</sup> Department of Crop Physiology, TNAU, Coimbatore-641003, Tamil Nadu, India. <sup>b</sup> Department of Plant Breeding and Genetics, TNAU, Coimbatore 641003, India. <sup>c</sup> Tamil Nadu Rice Research Institute, Aduthurai 612101, India.

#### Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/IJPSS/2022/v34i232530

**Open Peer Review History:** 

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/93410

Original Research Article

Received 07 September 2022 Accepted 12 November 2022 Published 14 November 2022

#### ABSTRACT

Salt stress is a common abiotic stress that significantly restricts crop development and productivity. Physiological alterations in response to salt stress were assessed for fourteen rice varieties during the panicle initiation stage at 120mM NaCl. Statistical analysis program (SPSS 15.0) was used to carry out the study. Under salt stress, all the rice varieties were assessed manifested a reduction in chlorophyll accumulation, stable chlorophylls, membranes and hydration status. On the other hand, all the varieties showed an increase in proline, hydrogen peroxide, and superoxide dismutase activity. It is noteworthy that the rice varieties *surakuruvai, kaivarasamba, mallam punchai,* and *mappillai samba* had better levels of salt tolerance than the salt-sensitive ones due to increased SOD activity, proline accumulation, relative water content, chlorophyll, and membrane stability index. The ability to tolerate salt during the reproductive stage under field conditions will be further investigated using these varieties.

Keywords: CSI proline; RWC; SOD; salt tolerance rice.

#### **1. INTRODUCTION**

"Rice is the most important global food crops that providing food for more than half of the world's

population. However, rice productivity in several areas is affected by salinity stress due to the buildup of underground salt and is exacerbated

<sup>\*</sup>Assistant Professor;

Associate Professor;

<sup>&</sup>lt;sup>†</sup> Director;

<sup>\*</sup>Corresponding author: E-mail: vanitha.k@tnau.ac.in;

by salt mining, deforestation, and irrigation. Nearly 1 billion hectares of land on Earth are affected by salinity, which damages 900 million ha of land, or almost 20% of all land on Earth. Additionally, about half of all irrigated arable land on Earth is affected by salinity" [1]. The most pervasive issue with soil toxicity in nations that grow rice is soil salinity. Because appropriate agricultural land is scarce, boosting rice's salinity resistance is essential for further expanding the rice-growing region.

"However, damage can also ensue with the results of excessive reactive oxygen species (ROS) such as superoxide radicals  $(O_2)$ , hydrogen peroxide  $(H_2O_2)$  and hydroxyl radicals (OH) which are produced at a fast rate accumulated in a plant tissue as a result of ion imbalance and hyperosmotic stresses. As a result. ROS accumulation causes lipid oxidation which has a detrimental effect on cellular metabolism and physiology thus adversely destructs the membrane integrity" (Munns et al. 2006). "Several plants have developed mechanisms to regulate the synthesis and accumulation of compatible solutes like proline alycine betaine that serve and as osmoprotectants and are essential for plants to adapt to osmotic stress by stabilizing the tertiary structure of proteins, in addition to ion homeostasis strategies" [2].

"Salinity-induced yield reduction of rice is alarming for the food security of the ever-growing population of the world, especially in Asia, because 90% of the world's rice is produced and consumed in Asia and more than 3 billion Asian intakes their 50-80 % daily calorie from rice" [3]. Keeping this in view, the present study was conducted to explore growth and physiological changes in rice varieties subjected to salinity stress differing in their level of salt tolerance.

#### 2. MATERIALS AND METHODS

# 2.1 Experimental Site, Plant Material and Salt Stress

This study was conducted in pot culture at the glasshouse of Tamil Nadu Rice Research Aduthurai, India. Fourteen Institute, rice genotypes (local types) were collected from farmer's fields over Tamil Nadu and screened for salt tolerance levels the their at early reproductive stage which is the panicle initiation stage. Rice seeds were surface sterilized, and directly sewn into pots (15 cm in height and 30

cm diameter). Ten kg of soil was collected from paddy fields and mixed with river sand and FYM in a 4:1:1 ratio. This experiment was laid out in a complete randomized block design with four replications. Best genotypes from the previous hydroponic study were used for pot culture (Preliminary study already completed). Pokkali, a well-known salt-tolerant genotype was used as a standard tolerant check and IR64 as a susceptible check. Salt solutions were prepared by dissolving NaCl salt in water with a concentration of 6 EC (60 mM NaCl) and subsequently rose to 12 dS/m (120 mM NaCl). Then pots were irrigated with saltwater and salinity levels were closely monitored for each treatment. Seedlings of each rice variety were subjected to salinity stress at 120 mM NaCl for 15 days during panicle initiation (50 to 65 DAS).

Sampling was performed at the end of the experiments and physiological changes were recorded. Chlorophyll content was measured by the method of Arnon [4]. Relative water content (RWC) was calculated using the formula proposed by Weatherley [5]. Proline content was estimated by the modified procedure of Bates et al. [6] and expressed as  $\mu g g^{-1}$  tissue FW. Chlorophyll Stability Index (CSI) in the leaf was estimated using the method of Koleyoreas [7]. Membrane Stability Index (MSI) was determined by the estimation of electrolyte leakage in leaf samples by using the method proposed by Pinhero and Fletcher [8]. The content of H<sub>2</sub>O<sub>2</sub> was measured by the method of Velikova et al. [9]. The SOD activity was assayed by the method of Beauchamp and Fridovich [10].

The pot culture experiments were arranged in a completely randomized design. The data were subjected to one-way analysis of variance (ANOVA) as suggested by Gomez and Gomez, 1984, and to mean separation with the Fisher's Least Significant Differences (LSD) test with P<0.05, using the statistical analysis program (SPSS 15.0).

#### 3. RESULTS & DISCUSSION

Salinity can cause negative effects on plant growth and development. The adaptive behaviour of rice varieties under a salt stress environment is discussed hereunder through various physiological and biochemical aspects. Chlorophyll pigments play a vital role in crop productivity because these pigments are highly responsible for photosynthesis in plants. In our study, the chlorophyll a, b, and total contents generally declined under salt stress which is in line with Wang et al. [11]. Accordingly, tolerant genotypes had higher chlorophyll content than susceptible ones. The genotypes namely mallam punchai and surakuruvai recorded higher chlorophyll a, b and total content under salt stress conditions. This might be due to better protection of salt-induced chlorophyll loss in rice cultivars with higher salt tolerance was also observed in earlier reports by Khan and Abdallah [12]. "The lowest was recorded in chinnapunchai, uppumilagai and IR64 under salt stress conditions than the rest of the genotypes tested. Salt stress reduced the amount of chlorophyll in the leaves by degrading or inhibiting the synthesis of chlorophyll" [13]. "The increased rate of chlorophyllase enzyme activity (enzymes degrading chlorophyll) is favoured under high salt conditions. This might be one of the important factors for the reduction of photosynthesis under salt stress. Hence, variation in the chlorophyll content can be used as a stress indicator" [14], "because chlorophyll content decreased in sensitive crop plants under salt stress conditions" [13].

RWC is the indicator of the water status of the plant. Salt stress significantly affects the water status of the plants. All the genotypes maintained good water status under well-watered conditions. A higher leaf RWC value of 83.5, 82.5, 81.5 and 81.4 percent were evident with the genotypes namely kattaikar, kaivarasamba, kallundaikar and surakuruvai respectively which was on par with the tolerant check Pokkali, which were also significantly superior to the rest of the genotypes under salt stress condition. "It seems that these genotypes were able to maintain the relatively high turgidity required for leaf function. Since sensitive genotypes usually transfer larger amounts of Na from roots to shoots. this could result in higher osmotic potential in their roots and less water uptake from the saline soil solution. Sensitive genotypes are have also known to а less stomatal function when subjected to salt stress, resulting in higher transpiration and greater water loss, both of which could be reflected in lower values of leaf RWC and consequent cellular dehydration" [15].

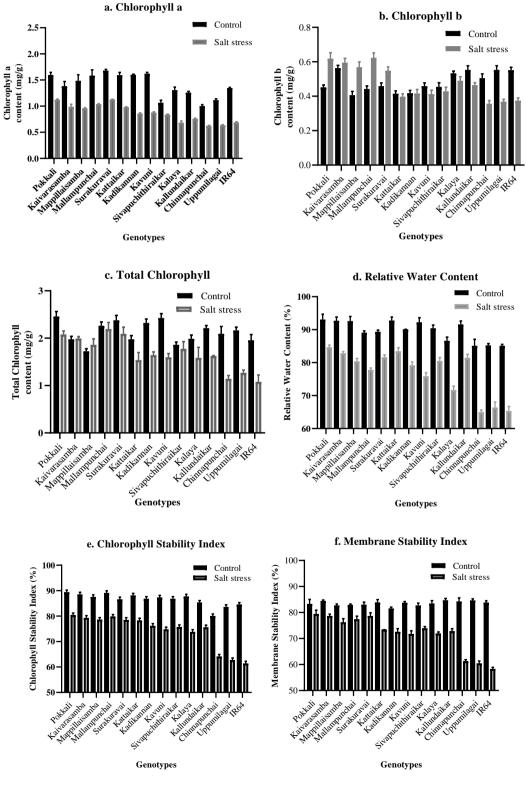
The CSI is an indication of the stress tolerance capacity of plants. In all the investigated genotypes, the CSI and MSI percentage decreased in genotypes under salt stress conditions. But there were no significant differences between the genotypes under control

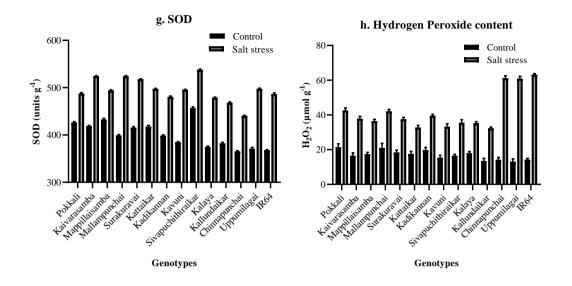
conditions (Table 2). Among the genotypes, kaivarasamba, mappillai samba, mallam punchai, surakuruvai and kattai kar had significantly higher CSI percentages (79.36, 78.65, 79.94, 78.61 and 78.36% respectively) under salt stress conditions along with the tolerant check (80.50%). A high CSI value means that the stress did not have much effect on the chlorophyll content of plants and also helps the plants to withstand stress conditions through better availability of chlorophyll. This leads to an increased photosynthetic rate, more dry matter production and higher productivity. We also observed significant positive correlations of RWC with CSI (r = 0.842, P<0.01), MSI (r = 0.660, P<0.05),  $H_2O_2$  (r = 0.709, P < 0.01), proline (r = 0.774, P<0.01), chlorophyll a (r = 0.819, P<0.01), chlorophyll b (r = 0.558, P<0.05) and total chlorophyll (r = 0.714, P<0.01) furnished in Table 1.

"MSI is an indicative of salt tolerance as it measures the extent of cell membrane injury under stress, as observed previously for salt stress" [16]. Like RWC, MSI was also significantly higher in tolerant genotypes viz., kaivarasamba (79.60%), mappillai samba mallam punchai (77.45%) (76.23%). and surakuruvai (78.60%). We also observed significant positive correlations of MSI with SOD (r = 0.56, P<0.05), proline (r = 0.778, P<0.01),  $H_2O_2$  (r = 0.789, P < 0.01), chlorophyll a (r = 0.897, P<0.01), chlorophyll b (r = 0.830, P<0.01) and total chlorophyll (r = 0.893, P<0.01) (Table 2).

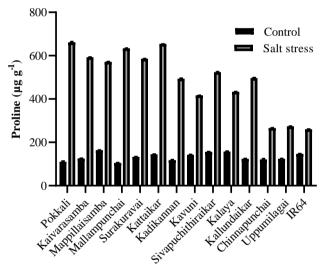
The result of the present study showed that the amount of  $H_2O_2$  varied among different genotypes under salt stress conditions. Salt ROS production stress increases which automatically activates the antioxidant enzymes in the tolerant plants. Higher amount of  $H_2O_2$ accumulated in the susceptible genotypes namely china punchai (61.14  $\mu$ mol g<sup>-1</sup>), uppu milagai (60.86  $\mu$ mol g<sup>-1</sup>) and *IR64* (63.21  $\mu$ mol g<sup>-1</sup>) <sup>1</sup>). seems that NaCl-induced  $H_2O_2$ It accumulation reduces plant growth, development and productivity [17]. "The removal of the free oxygen radicals is an important mechanism of salt tolerance in plants" [18]. In this present study, the salinity stress affected the activity of the antioxidant system which is in line with Wi et [19]. Since the H<sub>2</sub>O<sub>2</sub> content varied al. significantly among the varieties, the activity of major H<sub>2</sub>O<sub>2</sub> scavenging enzyme SOD was variable in these genotypes under the salt stress conditions [20-22]. Interestingly, salt stress

increased the SOD activity in all the genotypes tested than control. Among the genotypes, *kaivarasamba, mappillai samba, mallam punchai, surakuruvai, kadi kannan* and *sivapuchithirai kar* recorded significantly higher SOD activity under salt stress conditions. Thus, it seems that the genotypes had an efficient enzymatic detoxification system for  $H_2O_2$  scavenging. "The higher activity of SOD was also observed in other salt-tolerant plants" [23,24].









Genotypes

## Fig. 1. Comparison of physiological parameters in rice genotypes exposed to salt stress condition

1a. Chlorophyll a content (mg g<sup>-1</sup>), 1b. Chlorophyll b content (mg g<sup>-1</sup>), 1c. Total Chlorophyll content (mg g<sup>-1</sup>), 1d. Relative Water Content (%), 1e. Chlorophyll Stability Index (%), 1f. Membrane Stability Index (%), 1g. Super Oxide Dismutase (SOD: units g<sup>-1</sup>), 1h. H<sub>2</sub>O<sub>2</sub> content (μmol g<sup>-1</sup>), 1i. Proline content (μg g<sup>-1</sup>)

Proline acts as a compatible solute which seems to have diverse adaptive roles including stabilization of proteins and stabilization of membrane and sub-cellular structures [25], protecting cellular functions by scavenging reactive oxygen species [26], the storage form of carbon to provide the energy needed for recovery [27] and acting as a signal molecule controlling reproductive development [28]. Igarashi et al. [29] suggested that proline accumulation was related to the degree of salt tolerance. "The accumulation of high proline content in the rice cultivars under salt stress was able to maintain a higher green leaf area" [30]. Accordingly, in the present study, the proline content was increased in all the rice genotypes under salt stress conditions than in the control.

Genotypes	RWC	CSI	MSI	SOD	$H_2O_2$	Proline	Chl a	Chl b	ТС
Pokkali	0.91	0.89	0.92	1.14	2.00	5.08	0.70	1.36	0.83
Kaivarasamba	0.89	0.91	0.88	1.25	2.27	3.95	0.75	1.05	0.99
Mappillai samba	0.87	0.90	0.91	1.14	2.10	2.88	0.64	1.41	1.08
Mallam Punchai	0.87	0.90	0.93	1.31	2.01	6.05	0.65	1.41	0.98
Surakuravai	0.91	0.91	0.94	1.25	2.05	4.41	0.67	1.20	0.89
Kattai kar	0.90	0.89	0.88	1.19	1.88	4.53	0.61	0.94	0.79
Kadi kannan	0.87	0.88	0.89	1.21	1.98	4.19	0.53	0.98	0.71
Kavuni	0.83	0.86	0.86	1.29	2.15	2.93	0.54	0.89	0.66
Sivapuchithiraikar	0.89	0.87	0.89	1.18	2.16	3.36	0.79	0.94	0.94
Kalaya	0.83	0.84	0.86	1.28	1.41	2.76	0.52	0.91	0.79
Kallundai kar	0.89	0.89	0.85	1.22	1.64	4.02	0.60	0.85	0.73
Chinna punchai	0.84	0.85	0.85	1.21	1.50	2.21	0.61	0.71	0.54
Uppumilagai	0.83	0.85	0.84	1.34	1.60	2.22	0.57	0.67	0.59
IR64	0.85	0.81	0.85	1.32	1.64	1.78	0.51	0.68	0.54

Table 1. Salt tolerance index of physiological parameters in rice genotypes under salt stress

<sup>a</sup> Salt tolerance index was defined as the observations under salt stress divided by the means of the controls

 
 Table 2. Pearson correlation coefficients among physiological parameters from rice genotypes under salt stress

Parameters	RWC	CSI	MSI	SOD	H <sub>2</sub> O <sub>2</sub>	Proline	Chl a	Chl b	Total Chl
RWC	1								
CSI	0.842**	1							
MSI	0.660*	0.786**	1						
SOD	0.424	0.604*	0.560*	1					
$H_2O_2$	0.709**	0.842**	0.789**	0.599*	1				
Proline	0.825**	0.889**	0.758**	0.521*	0.756**	1			
Chl a	0.819**	0.888**	0.897**	0.557*	0.895**	0.890**	1		
Chl b	0.558*	0.796**	0.830**	0.441	0.697**	0.509	0.756**	1	
тс	0.714**	0.928**	0.893**	0.626*	0.850**	0.754**	0.873**	0.887**	1

\*\* Correlation is significant at the 0.01 level (2-tailed), \* Correlation is significant at the 0.05 level (2-tailed)

#### 4. CONCLUSION

The results of this study showed that the rice genotypes namely *kaivarasamba, mappillai samba, mallam punchai* and *surakuruvai* possessed higher degrees of salt tolerance by enhanced activity of physiological traits such as RWC, CSI, MSI, Proline and SOD activity. These genotypes could be further investigated at the reproductive stage salt tolerance ability under salt affected soil conditions.

#### **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

#### REFERENCES

1. Velmurugan A, Swarnam P, Subramani T, Meena B, Kaledhonkar MJ. Water demand and salinity. In: Desalination-challenges and opportunities. 2020;1-11.

- Munns R, Tester M. Mechanisms of salinity tolerance. Annu. Rev. Plant Biol. 2008; 59:651–681.
- 3. Khush G. What it will take to feed 5.0 billion rice consumers in 2030. Plant Mol. Biol. 2005;59:1–6.
- 4. Arnon DI. Copper enzymes in isolated chloroplast and polyphenol oxidase in Beta Vulgaris. Plant Physiol. 1949;(24)1:1-15.
- 5. Weatherely PE. Studies in the water relations of the cotton plant. I. The field measurements of water deficits in leaves. New Phytol. 1950;49: 81-87.
- Bates LS, Waldren RP, Teare ID. Rapid determination of free proline for water stress studies. Plant Soil. 1973;39(1):205-207.
- Koleyoreas SA. A new method for determining drought resistance. Plant Physiol. 1958;33:1-22.

- Pinhero RG, Fletcher RA. Paclobutrazol and ancymidol protect corn seedlings from high and low temperature stresses. Plant Growth Regul. 1994;15(1): 47-53.
- Velikova V, Yordanov I, Edreva A. Oxidative stress and some antioxidant systems in acid rain-treated bean plants: Protective role of exogenous polyamines. Plant Sci. 2000;151: 59-66.
- 10. Beauchamp C, Fridovich I. SOD improved assays and an assay applicable to acrylamide gel. Anal Biochem. 1971;44(1):276-287.
- Wang L, Showalter A, Ungar I. Effect of salinity on growth, ion content, and cell wall chemistry in *Atriplex prostrata* (Chenopodiaceae). Am. J. Bot. 1997;84(9):1247.
- Khan MA, Abdallah Z. Salinity-sodicity induced changes in reproductive physiology of rice (*Oryza sativa*) under dense soil conditions. Environ. Exp. Bot. 2003;49(2):145-157.
- 13. Ashraf M, Harris P. Photosynthesis under stressful environments: an overview. Photosynthetica. 2013;51:163-190.
- 14. Naumann JC, Young DR, Anderson JE. Leaf chlorophyll fluorescence, reflectance, and physiological response to freshwater and saltwater flooding in the evergreen shrub, *Myrica cerifera*. Environ Exp Bot. 2008;63:402-409.
- 15. Qin J, Dong WY, He KN, Yu Y, Tan GD, Han L, Dong M, Zhang YY, Zhang D, Li AZ, Wang ZL. NaCl salinity-induced changes in water status, ion contents and photosynthetic properties of *Shepherdia argentea* (Pursh) Nutt. seedlings. Plant Soil Environ. 2010;56:325-332.
- Bhattacharjee S, Mukherjee AK. Ethylene evolution and membrane lipid peroxidation as indicators of salt injury in leaf tissues of Amaranthus lividus seedlings. Ind. J. Exptl. Biol. 1996; 34:279-281.
- Vaidyanathan H, Sivakumar P, Chakrabarsty R, Thomas G. Scavenging of reactive oxygen species in NaCI-stressed rice (*Oryza sativa* L.) differential response in salt-tolerant and sensitive varieties. Plant Sci. 2003;165:1411-1418.
- Motohashi T, Nagamiya K, Prodhan SH. Production of salt stress tolerant rice by over expression of the catalase gene, katE, derived from Escherichia coli. Asia Pac. J. Mol. Biol. Biotechnol. 2010;18:37-41.

- 19. Wi SG, Chung BY, Kim JH, Lee KS, Kim JS. Deposition pattern of hydrogen peroxide in the leaf sheaths of rice under salt stress. Biol Plant. 2006;50:469-472.
- 20. Munns R. Comparative physiology of salt and water stress. Plant Cell Environ. 2002;25(2):239-250.
- 21. Munn AJ, Snelling EP, Taggart DA, Clauss M. Scaling at different ontogenetic stages: Gastrointestinal tract contents of a marsupial foregut fermenter, the western grey kangaroo Macropus fuliginosus melanops. Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology. 2022 Feb 1;264:111100.
- 22. Tabassum R, Tahjib-UI-Arif Μ, Hasanuzzaman M. Sohad AA. Islam MS. Shafi SS, Islam MM, Hassan L. Screening salt-tolerant rice at the seedling and reproductive stages: An effective and reliable approach. Environmental and Experimental Botany. 2021 Dec 1:192:104629.
- 23. Sekmen AH, Türkan I, Takio S. Differential responses of antioxidative enzymes and lipid peroxidation to salt stress in salt-tolerant Plantago maritima and salt-sensitive Plantago media. Physiol Plant. 2007;131(3):399-411.
- 24. Sairam RK, Rao KV, Srivastava G. Differential response of wheat genotypes to long term salinity stress in relation to oxidative stress, antioxidant activity and osmolyte concentration. Plant Sci. 2002;163:1037-1046.
- Van Rensburg L, Krüger GHJ, Krüger H. Proline accumulation as drought-tolerance selection criterion: its relationship to membrane integrity and chloroplast ultra structure in *Nicotiana tabacum* L. J Plant Physiol. 1993;141:188-194.
- 26. Smirnoff N, Cumbes QJ. Hydroxyl radical scavenging activity of compatible solutes. Phytochemistry. 1989;28(4):1057-1060.
- 27. Hare PD, Cress WA. Metabolic implications of stress- induced proline accumulation in plants. Plant Growth Regul. 1997;21:79-102.
- Mattioli R, Marchese D, D'Angeli S, Altamura MM, Costantino P, Trovato M. Modulation of intracellular proline levels affects flowering time and inflorescence architecture in Arabidopsis. Plant Mol. Biol. 2008;66(3):277-288.
- 29. Igarashi Y, Yoshiba Y, Sanada Y, Yamaguchi-Shinozaki K, Wada K, Shinozaki K. Characterization of the gene

for  $\Delta^1$ -pyrroline-5-carboxylate synthetase and correlation between the expression of the gene and the salt tolerance in *Oryza sativa* L. Plant Mol. Biol. 1997;33(5): 857 -865.  Kordrostami M, Rabiei B, Kumleh HH. Biochemical, physiological and molecular evaluation of rice cultivars differing in salt tolerance at the seedling stage. Physiol. Mol. Biol. Plants. 2017; 23(3):529-544.

© 2022 Vanitha et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle5.com/review-history/93410