



## Status of Nitrogen Pools Under Important Cropping Systems in Inceptisols and Vertisols of Northern Telangana, India

V. Shalini <sup>a#</sup>, A. Krishna Chaithanya <sup>b†</sup>, K. Chandrashaker <sup>b‡</sup>  
and CH. Aruna Kumari <sup>c‡</sup>

<sup>a</sup> Department of Soil Science and Agricultural Chemistry, College of Agriculture, Rajendranagar, 500030, India.

<sup>b</sup> Regional Agricultural Research Station, PJTSAU, Jagtial -505529, India.

<sup>c</sup> Department of Crop Physiology, College of Agriculture, Rajendranagar- 500030, Professor Jayashankar Telangana State Agricultural University, India.

### Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

### Article Information

DOI: 10.9734/IJPSS/2022/v34i232538

### Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/93040>

Original Research Article

Received 03 September 2022

Accepted 01 November 2022

Published 17 November 2022

### ABSTRACT

Vertisols have showed 11.58, 26.92 and 19.80 % higher amount of available nitrogen, 20.42, 36.65 and 16.72 % higher amount of total nitrogen in 0-15, 15-30 and 30-45 cm depths, respectively over inceptisols. CS<sub>1</sub> has maintained higher amount of available nitrogen content (237 kg ha<sup>-1</sup>) followed by CS<sub>2</sub> (219 kg ha<sup>-1</sup>) > CS<sub>4</sub> (189 kg ha<sup>-1</sup>) > CS<sub>3</sub> (184 kg ha<sup>-1</sup>) at surface soil (0-15 cm). In vertisols, ammonical nitrogen contributed 48.72 percent to available N, whereas in inceptisols it was 44.46%. Nitrate nitrogen content was recorded significantly higher under vertisols at 0-15 and 15-30 cm soil depths. At 30-45 cm depth inceptisols recorded significantly higher values. However, NO<sub>3</sub>-N contributed 32.52 percent towards available N in inceptisols, whereas the share was 30.33 percent under vertisols. In the soil profile, percent contribution of ammonical nitrogen to available N followed as CS<sub>1</sub> > CS<sub>4</sub> > CS<sub>2</sub> > CS<sub>3</sub> with the values 53.13, 48.26, 44.49 and 41.49 %. On the other hand, percent contribution of NO<sub>3</sub> -N towards available N followed different order as CS<sub>4</sub> (40.08%) > CS<sub>3</sub> (37.42%) > CS<sub>2</sub> (32.44%) > CS<sub>1</sub> (20.92%).

# Ph. D Scholar,

† Scientist at RARS,

‡ Assistant Professor,

\*Corresponding author: E-mail: shaliniagbsc@gmail.com;

**Keywords:** Vertisols; inceptisols; cropping systems; ammonical-N; nitrate-N.

## 1. INTRODUCTION

Nitrogen (N) is assumed to be the most yield-limiting nutrient element for crop production throughout the globe and is applied in the largest quantity for most of the annual crops [1]. Sustainability of an agricultural production system that depends highly on the soil reserve to meet the N requirements cannot be effective for long in producing high yields of crops [2]. Except for legumes, which have the ability to fix their own N, N must be supplied externally to plants for growth. It is usually added as a fertilizer and is required for all types of soils [3]. To increase crop yields, growers worldwide apply over 80 million metric tons of nitrogen fertilizers per year [4]. Use of inorganic N fertilizers has had its most substantial beneficial effect on human health by increasing the yield of field crops and nutritional quality of foods needed to meet dietary requirements and food preferences for growing world populations [5,6]. Ridley and Hedlin [7] concluded that increased use of N fertilizer has had the most dramatic influence on increasing crop yields since the 1950s, in combination with disease-resistant cultivars to a lesser effect. Similarly, Camara et al. [8] reported that historically, few if any technologies have increased winter wheat yield in the United States more than N fertilization. The main reasons for N deficiency are high-quantity uptake by crop plants compared to other macronutrients (except K in some crops such as rice), also in grains or seeds, and its loss by leaching, denitrification, volatilization, soil erosion, and surface runoff. In addition, N is immobilized by soil microbes and undecomposed plant residues, which may cause temporary deficiency. Nitrogen loss in the form of  $\text{NH}_3$  by plant canopy has been reported [9]. Furthermore, in intensive cropping systems, where no-tillage system is adopted, depletion or loss of organic matter has been reported [10], which may result in N deficiency in crop plants. Use of low rates for high-yielding modern crop cultivars, especially by farmers in developing countries, is another cause of N deficiency [11]. In developing countries, intensive agricultural production systems have increased the use of N fertilizer in efforts to produce and sustain high crop yields [11]. Consequently, N losses into the environment have also increased (Schmied et al. 2000). Even with the continuing research on N management, average worldwide N use

efficiencies (NUE) are reported to be around 50% [12,13], and N recovery efficiency for cereal production (rice, wheat, sorghum, millet, barley, maize, oat, and rye) is approximately 33% [14]. Understanding the effect of cropping systems on the transformation of organic N into different forms is a prerequisite for managing N inputs in a given soil. The present study was undertaken to quantify changes in soil carbon and nitrogen fractions and microbial process of C&N transformation in soil and their interrelationships under continuous cropping systems with differential nutrient management practices.

## 2. MATERIALS AND METHODS

Soil samples were collected from four cropping systems in Inceptisol and Vertisol at three depths. Sampling sites detailed description are given in Table 1.

### 2.1 Available Nitrogen

Available nitrogen content of soil was determined by using hot alkaline potassium permanganate (0.32%) for oxidative hydrolysis of the soil organic matter and liberated ammonia was absorbed and condensed in boric acid and titrated against standard 0.02 N  $\text{H}_2\text{SO}_4$  following the method as proposed by Subbiah and Asija [15].

### 2.2 Ammonical Nitrogen

Soil sample was extracted with 2 M KCL and filtered. Then the filtrate was steam distilled with 2.5% NaOH in the presence of 0.2 g MgO. The distillate was collected in 4% boric acid containing mixed indicator and was titrated with standard sulphuric acid (0.02 N) and expressed in  $\text{mg kg}^{-1}$  (Bremner, 1965).

### 2.3 Nitrate Nitrogen

Soil sample was extracted with 2 M KCL for an hour and filtered. Then the filtrate was steam distilled with 2.5% NaOH in the presence of 0.2 g Devarda's alloy to obtain  $\text{NO}_3\text{N}$ . the distillate was collected in 4% boric acid containing mixed indicator and was titrated with standard sulphuric acid (0.02 N) and expressed in  $\text{mg kg}^{-1}$  (Bremner, 1965).

**Table 1. Soil samples collected from the following experimental sites with GPS locations**

<b>Inceptisol</b>					
<b>Cropping system</b>	<b>Site-1</b>	<b>Site-2</b>	<b>Site-3</b>	<b>Site-4</b>	<b>Site-5</b>
<b>Rice-Rice</b>	Gangadahara	Ramadugu	Chennur	Nennal	Morthad
	18°32'32" 78°59'07"	18°39'27" 78°59'07"	18° 88'37" 79° 79'52 "	19° 06'75" 79° 58'76"	18° 86'53" 78° 43'61"
<b>Rice-Maize</b>	Gagadhara,	Gangadahar	Gutraipally	Chennur	Nennal
	18°36'30" 07°90'20"	18° 36'07" 07°90'15"	18° 84'51" 78° 98'48"	18° 88'44" 79° 79'53 "	19° 07'36" 79° 58'77 "
<b>Trmeric-Sesame</b>	Gangadhara	Ramadugu	Ramadugu	Chennur	Nennal
	18°36'07" 07°90'15"	18° 50'47" 07° 85'83"	18° 50'48 " 07° 85'82"	18° 88'14" 79° 30'53"	19° 08'36" 79° 59'58"
<b>Cotton Fallow</b>	Polasa	Padkal	Jakranpally	Kota armur	Morthad
	18° 84'44" 78° 95'44"	18° 69'75" 78° 27'55"	18° 71'55" 78° 26'54"	18° 79'31" 78° 32'66"	18° 81'86" 78° 41'07"
<b>Vertisol</b>					
<b>Rice-Rice</b>	Ragatlapalle	Gullapet	Mohanraopet	Anantharam	Chennur
	18° 22'38" 78° 41'11"	18° 86'14" 78° 95'86"	18° 81'30" 78° 76'46"	18° 85'34" 78° 97'63"	18° 73'11" 79° 80'51"
<b>Rice-Maize</b>	Upparmalyal	Anatharam	Thakalapally	Thakalapally	Chennur
	18° 32'38" 07° 85'90"	18° 85'35" 78° 97'73"	18° 85'41" 78° 97'30"	18° 85'43" 78° 97'36"	18° 88'41" 79° 77'53"
<b>Trmeric-Sesame</b>	Ragatlapalle	Chennur	Manchiryal	Chennur	Kalamadugu
	18° 22'34" 07° 84'10"	18° 73'38" 79° 80'40"	18° 95'20" 79° 45'36"	18° 73'32" 79° 80'36"	19° 08'62" 78° 95'22"
<b>Cotton-Fallow</b>	Palem	Palem,	Velpur	Thakallapally	Thakallapally
	18° 87'05" 78° 44'21"	18° 87'10" 78° 44'20"	18° 87'56" 78° 44'13 "	18° 85'28" 78° 97'68"	18° 85'28" 78° 97'77"

## 2.4 Total Nitrogen

Estimation of total nitrogen can be done by taking 2.0 g of soil in to 500 ml kjeldahl flask, the soil was swirled with 1g of salicylic acid and 20 ml of concentrated H<sub>2</sub>SO<sub>4</sub> for 30 minutes at room temperature. Then added 5g of sodium thiosulphate and 20 g of digestion mixture, allowed the contents for digestion and run the distillation unit by 40% NaOH, the released ammonia trapped in to 4% boric acid mixed indicator solution and titrated against 0.01 N H<sub>2</sub>SO<sub>4</sub> until bluish green colour turns pink, the used up H<sub>2</sub>SO<sub>4</sub> gives the titer value for calculating total nitrogen content in soil (Page et al. 1982).

## 3. RESULTS AND DISCUSSION

### 3.1 Nitrogen and its Pools

Nitrogen is the most limiting nutrient in crop production in India and the form of nitrogen available in the rhizosphere is considerably influenced by the presence of oxygen in the root zone. The available nitrogen, ammonical and

nitrate nitrogen concentration was significantly influenced by cropping systems under inceptisols and vertisols.

#### 3.1.1 Available-nitrogen

It is observed that all the soils comes under lower category of available nitrogen content, irrespective of soil depth it was ranging from 45 to 247 kg ha<sup>-1</sup>. Both the soil orders and cropping systems had significantly influenced available N content in soil (Table 1).

Vertisols have showed 11.58, 26.92 and 19.80% higher amount of available nitrogen content in 0-15, 15-30 and 30-45 cm depths, respectively over inceptisols. Irrespective of soil order, found an abrupt decline along soil depth, with middle (15-30 cm) and lower (30-45 cm) layers contained only 32.56 and 18.13% of total profile (0-45 cm) available nitrogen content. Available nitrogen content in soil primarily depends on organic residuals entering the soil, decomposition of organic matter [16] and on soil properties. Surface soil receives large amount of organic residuals like root biomass and exudates as active root zone of most of the crops limits to

0-20 cm soil depth. With increase in depth, residue return decreases, hence lower layers of soil showed low available nitrogen.

In cropping systems, CS<sub>1</sub> has maintained higher amount of available Nitrogen content (237 kg ha<sup>-1</sup>) followed by CS<sub>2</sub> (219 kg ha<sup>-1</sup>) > CS<sub>4</sub> (189 kg ha<sup>-1</sup>) > CS<sub>3</sub> (184 kg ha<sup>-1</sup>) at surface soil (0-15 cm). The same trend was observed in the sub surface soils also. CS<sub>1</sub> and CS<sub>2</sub> have shown significantly higher available nitrogen content in all the three soil depths over other cropping systems. Generally, high moisture of soil for a longer period of time contributes in accumulation of soil organic matter as well as soil nitrogen owing to soil anaerobic environment [17]. Anaerobic conditions were pronounced in CS<sub>1</sub> and CS<sub>2</sub> due to submergence might enhance soil available nitrogen by decreasing decomposition rate of soil organic matter. Moreover, greater below ground crop biomass produced under paddy crop [18,19] might helped CS<sub>1</sub> and CS<sub>2</sub> cropping systems in storing large amount of available soil nitrogen.

Interaction effect of soil orders and cropping systems were found to be non significant to influence available N in soils.

### 3.1.2 Total nitrogen, nitrate-nitrogen and ammonical-nitrogen

The amount of total nitrogen in soils was significantly higher in vertisols than inceptisols in all the three depths. Vertisols have shown 20.42, 36.65 and 16.72 % higher amount of TN in 0-15, 15-30 and 30-45 cm depths, respectively over inceptisols (Table 2). Relatively higher total nitrogen content in vertisol might be due to high clay content and lower values of TN in inceptisols may be associated with different parent material and its rate of disintegration [20]. Similar results were also reported by Das et al. [21] and Tabassum et al. [22]. Irrespective of soil order, found an abrupt decline in the amount of TN along soil depth, with middle (15-30 cm) and lower (30-45 cm) layers contained only 34.80 and 27.90% of total profile (0-45 cm).

Total Nitrogen (TN) content ranged from 799 to 1507 kg ha<sup>-1</sup> under different cropping systems. CS<sub>1</sub> has maintained significantly higher amount of TN (1493 kg ha<sup>-1</sup>) followed by CS<sub>2</sub> (1344 kg ha<sup>-1</sup>) > CS<sub>4</sub> (1253 kg ha<sup>-1</sup>) > CS<sub>3</sub> (1177 kg ha<sup>-1</sup>) at surface soil (0-15 cm). The same trend was observed in the sub surface soils also. CS<sub>1</sub> have shown significantly higher TN in all the three soil

depths over other cropping systems. Generally, high moisture of soil for a longer period of time contributes in accumulation of soil organic matter as well as soil nitrogen owing to soil anaerobic environment [17]. Anaerobic conditions were pronounced in CS<sub>1</sub> and CS<sub>2</sub> due to submergence might enhance soil available nitrogen by decreasing decomposition rate of soil organic matter. Moreover, greater below ground crop biomass produced under paddy crop [18,19] might helped CS<sub>1</sub> and CS<sub>2</sub> cropping systems in storing large amount of available soil nitrogen.

Interaction effect of soil orders and cropping systems were found to be non significant for total nitrogen in soils.

Soil types and cropping systems have significantly influenced the amount of ammonical and nitrate nitrogen content in soil. Ammonical nitrogen was recorded significantly higher under vertisols over inceptisols in all the three depths. With depth an abrupt decline in ammonical nitrogen content was observed. In vertisols, ammonical N contributed 48.72 percent to available N, whereas in inceptisols it was 44.46%. Amount of ammonia in soil primarily depends on mineralization rate and soil clay content. Dynamics of NH<sub>4</sub>-N adsorption and desorption in the dominant type of clay i.e. montmorillonite type [23], mineralization [24] and nitrification rates, which in turn is mediated by soil biomass [25-27] as well as the degree of K saturation in the intermediate layers of clay minerals [23] may be the reason for higher ammonical nitrogen under vertisols. Lower NH<sub>4</sub>-N under inceptisols was also reported by Dhamak et al. [28]. Such lower values may be due to lower clay content associated partly with different parent material and its rate of disintegration [20], besides lower amount of total N [28].

Nitrate nitrogen content was recorded significantly higher under vertisols with a value of 78.2 and 46.5 kg ha<sup>-1</sup> at 0-15 and 15-30 cm soil depths, respectively (Fig. 1). At 30-45 cm depth inceptisols recorded significantly higher values (23.6 kg ha<sup>-1</sup>) over vertisols (20.7 kg ha<sup>-1</sup>). However, NO<sub>3</sub>-N contributed 32.52 percent towards available N in inceptisols, whereas the share was 30.33 percent under vertisols. With depth, an abrupt decline of NO<sub>3</sub>-N was observed in both the soil types. NO<sub>3</sub> content in soil depends on nitrification rate soil aeration, moisture and type of microbial communities. Vertisols possess more bases and primary

minerals provide more favorable environment for diazotroph communities [29,30], hence recorded higher  $\text{NO}_3\text{-N}$ .

Cropping system also has significantly influenced both ammoniacal nitrogen and nitrate nitrogen content of soils.  $\text{CS}_1$  has recorded significantly higher amount of ammoniacal nitrogen in all the three depths with values of 131.7, 92.6 and 50.9  $\text{kg ha}^{-1}$  at 0-15, 15-30 and 30-45 cm depth, respectively as compared with other cropping systems.

Significantly lower values were recorded in  $\text{CS}_3$  at 0-15 and 15-30 cm depths, which was on par with  $\text{CS}_4$ . At 30-45 cm depth  $\text{CS}_4$  has shown significantly lower value, which was on par with  $\text{CS}_3$ . In the soil profile, percent contribution of ammoniacal nitrogen to available N followed as  $\text{CS}_1$  (53.13%) >  $\text{CS}_4$  (48.26%) >  $\text{CS}_2$  (44.49%) >  $\text{CS}_3$  (41.49%). In all the cropping systems  $\text{NH}_4\text{-N}$  values declined with depth.

Nitrate nitrogen values were recorded significantly higher under  $\text{CS}_2$  with values of 82.4 and 49.4 at 0-15 and 15-30 cm depth, which was on par with  $\text{CS}_4$  with values 79.8 and 43.3  $\text{kg ha}^{-1}$  (Fig. 1). At 30-45 cm depth  $\text{CS}_3$  has recorded higher values, on par with  $\text{CS}_2$ . In all the depths,  $\text{CS}_1$  has recorded significantly lower  $\text{NO}_3\text{-N}$  content. An abrupt decline was observed with depth in all the cropping systems.  $\text{NO}_3\text{-N}$  content

of soil in cropping system followed an order of  $\text{CS}_2 > \text{CS}_4 > \text{CS}_3 > \text{CS}_1$ . However, the percent contribution of  $\text{NO}_3\text{-N}$  towards available N followed different order as  $\text{CS}_4$  (40.08%) >  $\text{CS}_3$  (37.42%) >  $\text{CS}_2$  (32.44%) >  $\text{CS}_1$  (20.92%).

Results revealed that,  $\text{CS}_1$  has recorded significantly higher  $\text{NH}_4\text{-N}$  and lower  $\text{NO}_3\text{-N}$ , may be because of soil N and aeration interactions. Soil aeration was dependent on the soil moisture which affects the nitrogen release from organic and inorganic nitrogen sources [31]. In oxidized conditions thermodynamically stable form of N was nitrate ions, while under reduced or moderately oxidized conditions, ammonium ions will dominate [32]. Soils under  $\text{CS}_1$  were under submergence for 8-9 months in a year which causes anaerobic conditions, enhances redox potential. Such high redox potential restricts the conversion of ammoniacal N to nitrate N, as a result under  $\text{CS}_1$   $\text{NH}_4\text{-N}$  contributed lion share towards available N. Balance between ammoniacal and nitrate forms of nitrogen in soils also depends on the quality and quantity of nitrification, (function of availability of oxygen and microbial activity) and applied nitrogen [33]. The higher concentration of  $\text{NH}_4\text{-N}$  in  $\text{CS}_1$  may be due to reduced nitrification induced by lack of oxygen in soils because of continuous flooding during the crop growing season and reduced root growth resulted in lower nitrifying bacterial activity.

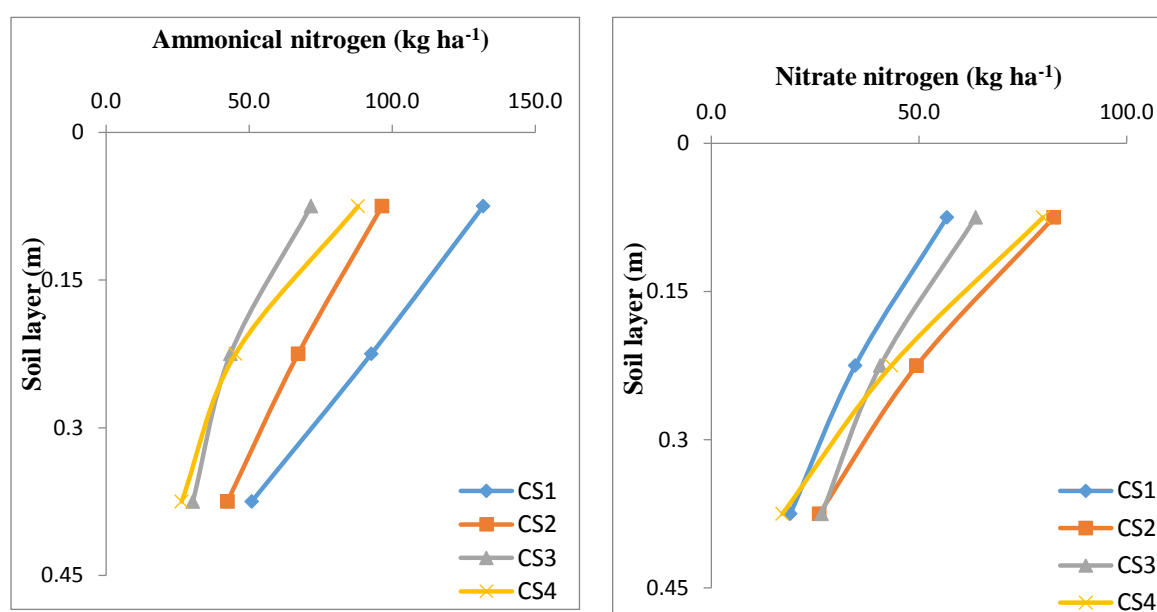
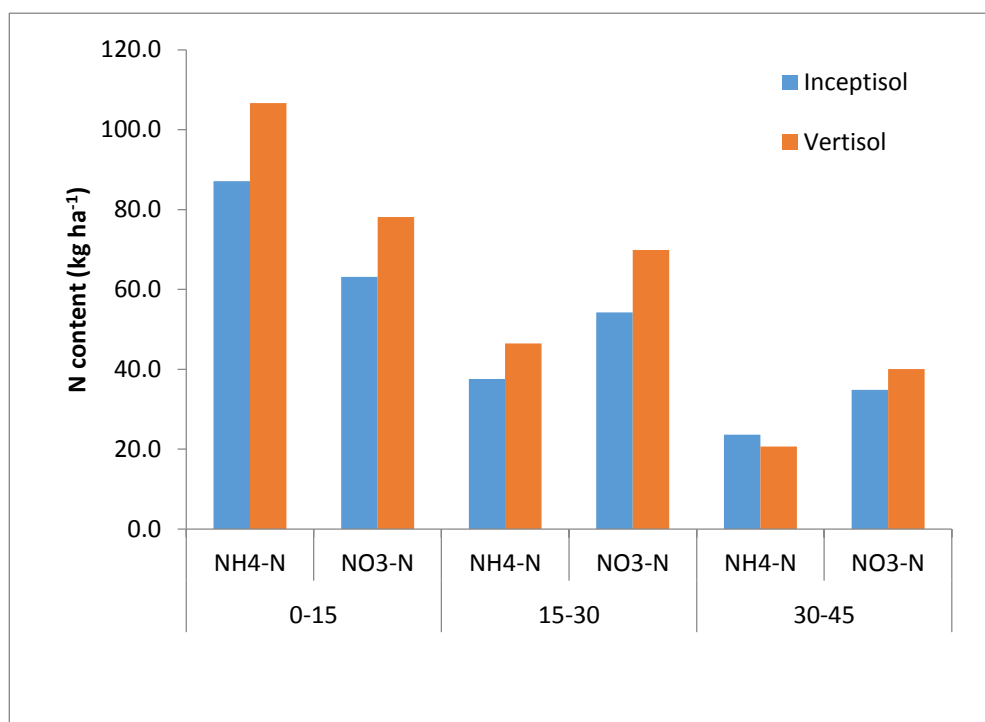


Fig. 1. Distribution of ammoniacal and nitrate nitrogen under different cropping systems along the depths

Table 2. Influence of soil type and cropping systems on soil nitrogen fractions and total nitrogen content of soils (kg ha<sup>-1</sup>)

Soil order	Available N (Kg ha <sup>-1</sup> )			Ammonical N (Kg ha <sup>-1</sup> )			Nitrate N (Kg ha <sup>-1</sup> )			Total N (Kg ha <sup>-1</sup> )		
	0-15	15-30	30-45	0-15	15-30	30-45	0-15	15-30	30-45	0-15	15-30	30-45
<b>S<sub>1</sub></b>	196	121	69	87.1	54.3	34.9	63.2	37.6	23.6	1195	1034	909
<b>S<sub>2</sub></b>	219	153	83	106.7	69.9	40.0	78.2	46.5	20.7	1439	1413	1061
<b>Sem</b>	7.64	5.85	4.5	3.5	2.0	1.5	2.7	1.9	1.0	45.2	44.65	34.36
<b>CD@5%</b>	22.14	16.94	13.04	10.1	5.9	4.2	7.7	5.5	2.8	130.93	129.35	99.54
<b>Cropping System</b>												
<b>CS<sub>1</sub></b>	237	175	100	131.7	92.6	50.9	56.7	34.6	19.0	1493	1487	1225
<b>CS<sub>2</sub></b>	219	172	84	96.4	67.1	42.4	82.4	49.4	26.0	1344	1227	922
<b>CS<sub>3</sub></b>	184	101	71	71.6	43.4	30.2	63.7	40.7	26.6	1177	1056	799
<b>CS<sub>4</sub></b>	189	99	50	88.0	45.1	26.3	79.8	43.3	17.1	1253	1125	994
<b>Sem</b>	10.81	8.27	6.37	4.9	2.9	2.1	3.8	2.7	1.4	63.92	63.15	48.59
<b>CD@5%</b>	31.31	23.95	18.44	14.3	8.4	6.0	10.9	7.8	3.9	185.16	182.96	140.76
<b>Interactions</b>												
<b>S<sub>1</sub>CS<sub>1</sub></b>	227	159	91	118.8	84.2	49.5	51.8	31.6	20.2	1279	1275	1078
<b>S<sub>1</sub>CS<sub>2</sub></b>	208	152	76	85.5	54.4	39.4	74.9	43.9	27.1	1225	1017	863
<b>S<sub>1</sub>CS<sub>3</sub></b>	173	86	66	69.1	39.2	28.1	58.3	38.0	27.5	1067	896	788
<b>S<sub>1</sub>CS<sub>4</sub></b>	176	86	45	75.1	39.2	22.4	67.7	36.9	19.8	1207	991	908
<b>S<sub>2</sub>CS<sub>1</sub></b>	247	192	108	144.7	101.1	52.2	61.6	37.7	17.9	1706	1739	1371
<b>S<sub>2</sub>CS<sub>2</sub></b>	231	192	93	107.3	79.8	45.4	90.0	55.0	24.9	1463	1437	982
<b>S<sub>2</sub>CS<sub>3</sub></b>	195	117	75	74.0	47.6	32.3	69.0	43.3	25.6	1287	1216	811
<b>S<sub>2</sub>CS<sub>4</sub></b>	202	112	55	100.8	51.1	30.2	92.0	49.8	14.4	1299	1260	1079
<b>Sem</b>	15.29	11.69	9	7.0	4.1	2.9	5.3	3.8	1.9	90.39	89.3	68.72
<b>CD@5%</b>	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
<b>CV</b>	16.49	19.11	26.42	16.1	14.7	17.5	16.8	19.1	19.5	15.35	16.32	15.6

*S<sub>1</sub>- Inceptisols, S<sub>2</sub>- Vertisols, CS<sub>1</sub>- Rice-Rice, CS<sub>2</sub>- Rice-Maize, CS<sub>3</sub>- Cotton –Fallow, CS<sub>4</sub>- Turmeric-Sesame, SE m: Standard error of mean, CD: Critical difference, CV: Critical Variance*



**Fig. 2. Depth wise allocation of ammonical and nitrate nitrogen content under inceptisols and Vertisols**

Higher NO<sub>3</sub>-N was observed in the treatment CS<sub>2</sub>, where huge amount of fertilizers were added to soil (for maize as it is an exhaustive crop) along with huge crop residue return to the soil (from rice crop residue). Under CS<sub>2</sub> cropping system alternate aerobic and anaerobic conditions prevails. These situation might have caused maximum nitrification which converts different forms of organic nitrogen and applied inorganic ammonical fertilizers to NO<sub>3</sub>-N. Similar results were also supported by Prasad et al. [34] Santhy et al. (1998) and Jain et al. [35].

Interaction of soil orders and cropping systems were found to be non significant for total, ammonical and nitrate nitrogen of soils [36].

#### 4. SUMMARY AND CONCLUSION

Vertisols have showed 11.58, 26.92 and 19.80% higher amount of available nitrogen content in 0-15, 15-30 and 30-45 cm depths, respectively over inceptisols. In cropping systems, CS<sub>1</sub> has maintained higher amount of available nitrogen content (237 kg ha<sup>-1</sup>) followed by CS<sub>2</sub> (219 kg ha<sup>-1</sup>) > CS<sub>4</sub> (189 kg ha<sup>-1</sup>) > CS<sub>3</sub> (184 kg ha<sup>-1</sup>) at surface soil (0-15 cm). The same trend was observed in the sub surface soils also. In vertisols, ammonical N contributed 48.72 percent

to available N, whereas in inceptisols it was 44.46%. However, NO<sub>3</sub>-N contributed 32.52 percent towards available N in inceptisols, whereas the share was 30.33 percent under vertisols. In the soil profile, percent contribution of ammonical nitrogen to available N followed as CS<sub>1</sub> (53.13%) > CS<sub>4</sub> (48.26%) > CS<sub>2</sub> (44.49%) > CS<sub>3</sub> (41.49%). However, the percent contribution of NO<sub>3</sub>-N towards available N followed different order as CS<sub>4</sub> (40.08%) > CS<sub>3</sub> (37.42%) > CS<sub>2</sub> (32.44%) > CS<sub>1</sub> (20.92%).

#### COMPETING INTERESTS

Authors have declared that no competing interests exist.

#### REFERENCES

1. Huber DM, Thompson IA. Nitrogen and plant disease. (eds.) Datnoff, L.E, Elmer, W.H and Huber, D.M. Mineral nutrition and plant disease. The American Phytopathological Society, St Paul, Minnesota. USA. 2007;55:31-44.
2. Stevenson FJ. Organic forms of soil nitrogen. F. J. Stevenson, (ed.) Nitrogen in agricultural soils. Agronomy Monograph No. 22, American Society of Agronomy,

- Crop Science Society of America, Soil Science Society of America, Madison, Wisconsin, USA. 1982;67–122.
3. Clark RB. Plant response to mineral element toxicity and deficiency. In: M. N. Christiansen and C. F. Lewis (eds). *Breeding Plants for Less Favorable Environments*. John Wiley and Sons, New York, NY. 1982;71–73.
  4. Epstein E, Bloom AJ. *Mineral nutrition of plants: principles and perspectives*. 2nd edition Sinauer Associates, Inc. Sunderland, Mass; 2005.
  5. Galloway JN, Cowling EB. Reactive nitrogen and the World: 200 years of change. *AMBIO: A Journal of the Human Environment*. 2002;31(2):64-71.
  6. Galloway JN, Cowling EB, Seitzinger SJ, Socolow R. Reactive nitrogen: Too much of a good thing?. *AMBIO: A Journal of the Human Environment*. 2002;31:60-63.
  7. Ridley AO, Hedlin RA. Crop yields and soil management on the Canadian prairies, past and present. *Canadian Journal of Soil Science*. 1980;60(3):393-402.
  8. Camara KM, Payne WA, Rasmussen PE. Long-term effects of tillage, nitrogen and rainfall on winter wheat yields in the Pacific Northwest. *Agronomy Journal*. 2003;95:828-835.
  9. Fageria NK, Baligar VC, Bailey BA. 2005. Role of cover crops in improving soil and row crop productivity. *Communications in Soil Science and Plant Analysis*. 36:2733- 2757.
  10. Johnson PM, Mayrand K, Paquin M, eds, *Governing global desertification. Linking environmental degradation, Poverty and Participation*. Aldershot: Ashgate; 2006.
  11. Fageria NK. Plant tissue test for determination of optimum concentration and uptake of nitrogen at different growth stages in low-land rice. *Communications in Soil Science and Plant Analysis*. 2003;34:259–270.
  12. Newbould P. The use of nitrogen fertilizer in agriculture. Where do we go practically and ecologically? *Plant Soil*. 1989;115:297–311.
  13. Collins AL, Stromqvist J, Davison PS, Lord EI. Appraisal of phosphorus and sediment transfer in three pilot areas identified for the Catchment Sensitive Farming initiative in England: application of the prototype PSYCHIC model. *Soil Use and Management*. 2007;23:117-132.
  14. Raun WR, Solie JB, Johnson GV, Stone ML, Whitney RW, Lees HL, Sembiring H, Phillips SB. Micro-variability in soil test, plant nutrient, and yield parameters in bermudagrass. *Soil Science and Society of American Journal*. 1998;62:683-690.
  15. Subbaiah BV, Asija GL. A rapid procedure for the estimation available N in the soils. *Current Science*. 1956;25:259.
  16. Yang W, Xia L, Zhu Z, Jiang L, Cheng X, An S. Shift in soil organic carbon and nitrogen pools in different reclaimed lands following intensive coastal reclamation on the coasts of eastern China. *Scientific Reports*. 2019;9:5921.
  17. Whitting GJ, Chanton JP. Greenhouse carbon balance of wetlands: methane emission versus carbon sequestration. *Tellus B*. 2001;53:521-528.
  18. Krishna Chaitanya A, Majumder SP, Badole S, Padhan D, Datta A, Mandal B, Srinivas CH. Pools of organic carbon in soils under a long-term rice–rice system with different organic amendments in hot, sub-humid India. *Carbon Management*. 2020;11 (4): 331-339
  19. Krishna Chaitanya A, Majumder SP, Padhan D, Badole S, Datta A, Mandal B, Kiran G. Carbon dynamics, potential and cost of Carbon sequestration in double rice cropping system in semi arid southern India. *Journal of Soil Science and Plant Nutrition*. 2018;18 (2), 418-434.
  20. Ghatol SG, Malewar GU. Influence of texture and organic matter on the physical properties of Marathwada soils. *Research Bulletin, Marathwada Agricultural University*. 1978;2:10-11.
  21. Das I, Ghosh K, Ray SC, Mukhopadhyay PK, Ghosh SK. Status and distribution of Sulphur vis-à-vis taxonomic classwise distribution of sulphur in selected soil series of Inceptisols in West Bengal. *Journal of the Indian Society of Soil Science*. 2006;41:776-777.
  22. Tabassum S, Sammy R, Muneshwar, S, Biswas AK. Changes in organic and inorganic forms of nitrogen in a typical haplustert under soybean- wheat system due to conjoint use of inorganic fertilizers and organic manures. *Journal of the Indian Society of Soil Science*. 2010;58:76-85.
  23. Nieder R, Benbi DK, Scherer HW. Fixation and defixation of ammonium in soils: a review. *Biology and Fertility of Soils*. 2010;47:1-14.



24. Villasenor D, Zagal E, Stolpe N, Hirzel J. Relationship between mineralized nitrogen during anaerobic incubations and residual effect of nitrogen fertilization in two rice paddy soils in Chile. *Chilean Journal of Agricultural Research*. 2015;75:98-104.
25. Janssens IA, Landkreijer H, Matteucci G, Kowalski AS, Buchmann N, Epron D, Pilegaard K, Kutsch W, et al. Productivity over-shadows temperature in determining soil and ecosystem respiration across European forests. *Global Change Biology*. 2001;7:269-278.
26. Sainz HR, Echeverría HE, Barbieri PA. Nitrogen balance as affected by application time and nitrogen fertilizer rate in irrigated no-tillage maize. *Agronomy Journal*. 2004;96:1622-1631.
27. Sahrawat K. Organic matter and mineralizable nitrogen relationships in wetland rice soils. *Communications in Soil Science and Plant Analysis*. 2006;37:787-796.
28. Dhamak A, Meshram N, Waikar S. Evaluation of nitrogen fractionation in relation to physicochemical properties of soil in Ambajogai Tahsil of Beed District. *IOSR Journal of Agriculture and Veterinary Science*. 2014;7:81-85.
29. Stone MM, Kan J, Plante AF. Parent material and vegetation influence bacterial community structure and nitrogen functional genes along deep tropical soil profiles at the Luquillo Critical Zone Observatory. *Soil Biology and Biochemistry*. 2015;80:273–282.
30. Pajares S, Escalante AE, Noguez AM, García-Oliva F, Martínez-Piedragil C, Cram SS, Eguarte LE, Souza V. Spatial heterogeneity of physicochemical properties explains differences in microbial composition in arid soils from Cuatro Ciénegas, Mexico. *Peer J*. 2016;4:24-59.
31. Agehara S, Warncke DD. Soil moisture and temperature effects on nitrogen release from organic nitrogen sources. *Soil Science Society of America Journal*. 2005; 69(6):1844-55.
32. Husson O. Redox potential (Eh) and pH as drivers of soil/plant/microorganism systems: A transdisciplinary overview pointing to integrative opportunities for agronomy. *Plant Soil*. 2013;362:389-417.
33. Sooksa-Nguan T, Yakubov B, Kozlovskyy VI, Barkume CM, Howe KJ, Thannhauser TW, Rutzke MA, Hart JJ, Kochian LV, Rea PA, Vatamaniuk OK. Drosophila ABC transporter, DmHMT-1, confers tolerance to cadmium. DmHMT-1 and its yeast homolog, SpHMT-1, are not essential for vacuolar phytochelatin sequestration. *Journal of Biological Chemistry*. 2009; 284(1):354-362.
34. Prasad P, George J, Masto RE, Rout TK, Ram LC, Selvi VA. Evaluation of microbial biomass and activity in different soils exposed to increasing level of arsenic pollution: A laboratory study. *Soil and Sediment Contamination*. 2013;22(5):483-497.
35. Jain P, et al. Cyclic AMP signaling pathway modulates susceptibility of candida species and *Saccharomyces cerevisiae* to antifungal azoles and other sterol biosynthesis inhibitors. *Antimicrobial Agents and Chemotherapy*. 2003;47(10): 3195-201.
36. Stone JA, BATTERY BR. Nine forages and the aggregation of a clay loam soil. *Canadian Journal of Soil Science*. 1989;69:165-169.

© 2022 Shalini et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:

<https://www.sdiarticle5.com/review-history/93040>