Journal of Advances in Microbiology



21(9): 39-49, 2021; Article no.JAMB.73467 ISSN: 2456-7116

Phytochemical Analysis and *In vivo* Evaluation of Individual Activity of Ethanolic Leaf Extracts of *Azadirachta indica, Senna occidentalis* and *Momordica balsamina* against *Plasmodium berghei* Infected Mice

Adam Musa Bature^{1*}, Karderam Bukar Dikwa¹, Abdullah Isyaku Alhaji² and Deboral Madi Dibal¹

¹Department of Biological Sciences, Nigerian Defence Academy, Kaduna, Nigeria. ²Department of Biotechnology, Nigerian Defence Academy, Kaduna, Nigeria.

Authors' contributions

This work was carried out in collaboration among all authors. Authors AMB and KBD designed the study. Author AMB performed the statistical analysis. Authors AMB, KBD and AIA wrote the protocol and wrote the first draft of the manuscript. Authors AMB, KBD and AIA managed the analyses of study. Author AMB managed the literature searches. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/JAMB/2021/v21i930382 <u>Editor(s):</u> (1) Dr. Ana Cláudia Correia Coelho, University of Trás-os-Montes and Alto Douro, Portugal. <u>Reviewers:</u> (1) Anil Kumar, India. (2) Diriba Leta Weleni, Arba Minch University, Ethiopia. Complete Peer review History: <u>https://www.sdiarticle4.com/review-history/73467</u>

> Received 05 July 2021 Accepted 05 September 2021 Published 16 September 2021

Original Research Article

ABSTRACT

Malaria is a major cause of morbidity and mortality in tropics and subtropics region with Nigeria accounting for the highest proportions in Africa. This is accompanied with emerging resistance to available drugs, posing it a public health concern. This study is aimed at determining the *invivo* activity of the individual ethanolic leaf extracts of the *Azadirachta indica*, *Senna occidentalis* and *Momordica balsamina*. The leaves of *A. indica*, *S. occidentalis* and *M. balsamina* were subjected to preliminary phytochemical screening. Ethanolic extracts of leaves of plants was carried out and *invivo* evaluation of the individual activity of extracts determined using standard procedures. 55 mice were randomly divided into 11 groups lettered A – K; positive group, negative group and 9

*Corresponding author: E-mail: adambature@nda.edu.ng;

extract groups. Results showed that *M. balsamina* had the highest yield of 7.6%, followed by *A. indica* with 6.5% and *S. occidentalis* with 5.7%. The preliminary phytochemical screening revealed the presences of alkaloids, flavonoids, saponins, steroids, phenolics and tannin in all plants. The comparison of the individual study groups showed that *Senna occidentalis* is more effective at 600mg/kg dosage and prolonged survival of the mice in its group in the study period. This plant possessed significant (P-value <0.05) antiplasmodial activity, thus lowered parasitaemia in infected mice.

Keywords: Antiplasmodial; Senna; Momordica; Azadirachta; parasitaemia.

1. INTRODUCTION

Plasmodium parasites, causative agents of Malaria have been a major health problem in many tropical regions of the world. An undoubtedly life-threatening disease prominent in the less developed and developing countries in tropical and subtropical regions including Africa, Asia and parts of Americans [1]. This disease has a mortality rate of more than one million children annually [2]. Globally, an estimated 216 million cases of the disease occurred in 91 countries making it a global concern with Nigeria accounting for the highest morbidity cases of 23%. In endemic areas of sub-sahara Africa, about 50 million pregnancies occurs every year and pregnancy associated malaria, another public health concern could lead to maternal and fetal morbidity leading to about 100, 000 deaths of infant annually [3]. Notably is the presence of numbers of available drugs to control the disease, but the development of resistant to these drugs has placed it as a major cause of death globally. One of the first effective antimalaria agent that was used was guinine isolated from the bark of cinchona plant that was later used to synthesize choloroguine and primaguine [4]. The resurgence of herbal remedies in recent times has been attributed to preference of consumers for natural therapies, a greater interest in alternative medicine and dissatisfaction with the result from orthodox pharmaceuticals as a result of ineffectiveness [5], thereby, the need for potential treatment candidates against malaria. Azadirachta indica is a member of the Mahogamy family with its name derived from 'azaddhirakt', a Persian word for 'noble tree' [6]. Its straight trunk measures bout 7-15m at maturity producing yellow ellopsidal drupes called neem fruits. The leaves are alternate, compound with its rachis length 15-25cm long. [7]. Senna occidentalis is an annual weed distributed widely in the tropical and subtrophical regions of the world [8] with height 60-150cm bearing ovate-lanceote ranging shaped compound leaves of 15-20 cm long [9],

Momordica balsamina is a trailer with its stem measuring 4-5 lenath mm. It is an annual/perennial climber plants usually 10-12inch tall and slighty hairy. The tendrils are simple and leaves bright green, palmate (5 - 7)lobed), waxy with lower surface paled more than the upper part. The plant bears red fruits with rows of cream short spines [10]. Although, the knowledge of its activity is known, comparing the invivo individual activity of the ethanolic extract of these plants would provide more essential information on their activity against Plasmodium berghei ANKA strain.

2. MATERIALS AND METHODS

2.1 Study Area

Adult mice weighing 20–30g of about 6–8 weeks old were purchased from the Veterinary Parasitology Laboratory, Ahmadu Bello University, Zaria and housed in aluminum cages containing wood shavings as beddings in the Biology Laboratory Section, Nigerian Defence Academy, Kaduna. The mice was acclimatized for 2 weeks where they were given pellets in small aluminum plates and drinking water through drinkers.

2.2 Collection, Authentication and Preparation of Plant Materials

Fresh leaves of A. indica, S. occidentalis and M. balsamina were collected from Kurmin Mashi and Zonkwa in Kaduna State. Plants were taxonomically authenticated at the department of Biological Sciences, Nigerian Defence Academy Kaduna. Leaves of plants were spread on a laboratory bench and allowed to air dry under normal room temperature. The dried leaves were sorted out to eliminate plant stems and branches and consequently coarsed into powder using mortar and pestle, followed by an electric mill. The powdered plant leaves were weighed, stored in a polythene bag and kept at room temperature until extraction.

The leaves of *A. indica, S. occidentalis* and *M. balsamina* were extracted using cold extraction. The leaves were soaked in distilled ethanol individually in 3 different glass jar. The solution was shaken at intervals to allow proper mixing in order to derive optimum extraction for 5 days, filtered using a filter paper. The filtrate was concentrated using water bath distillation process and the concentrate left open to air dry. The crude extract of *A. indica* (17g), *S. occidentalis* (16g), *M. balsamina* (20g) were kept in air-tight containers. The percentage yield of plant extracts were calculated using the method described by [11].

Percentage yield =
$$\frac{(W_2 - W_1) \times 100}{W_0}$$

Where W_2 is the weight of extract and container, W_1 is the weight of the container alone and W_0 is the weight of the initial dried leaves

2.4 Phytochemical Analysis

Phytochemical analysis of plants extracts were carried out using the method described by [12]. A small sample of the crude extracts of each of the plant was dissolved in 10 ml distilled ethanol. The solution was divided into various test tubes to test for the presence of the following photochemical:

2.4.1 Test for alkaloids

One cm³ of Hydrochloric acid (HCl) were added to the test fraction in a test tube. The solution was treated with Mayer, Wagner and Dragendorff reagent separately. A creamy-white (Mayer), reddish-brown (Wagner), and orange-brown (Dragendorff) precipitate indicates the presence of alkaloids.

2.4.2 Test for phenolics and tannins

Three drops of ferric chloride were added to the test extract solution in a test tube. A dirty green precipitate indicates a positive test.

2.4.3 Test for saponins

Three drops of olive oil were added to the test extract solution and the mixture was shaken vigorously. A stable emulsion formed indicates the presence of saponins.

2.4.4 Test for steroids

Three drops of H_2SO_4 and acetic anhydride were added to the test extract solution. Color changes from violet to blue or green indicates a positive test.

2.4.5 Test for Terpenoids

Three drops (2 ml) of chloroform were added to the test extract solution and evaporated on the water bath. it was boiled with 3ml of concentrated H_2SO_4 . A grey color formed shows the entity of terpenoids.

2.4.6 Test for flavonoids

Three drops (2 ml) of 2.0 % sodium hydroxide solution were added to the test extract. The formation of intense yellow color which becomes colorless on addition of 2 drops of dilute hydrochloric acid indicates the presence of flavonoids.

2.4.7 Test for reducing sugar

To test for the presence of reducing sugar in the extract, 5 - 6 drops each of Fehling solution A and B were added to the solution. The mixture were boiled and a brick red precipitate indicates of a positive test.

2.5 Parasite Strain

Plasmodium berghei ANKA strain was obtained from Veterinary Parasitology Laboratory in Ahmadu Bello University, Zaria where infected mice are being acclimatize. The parasite is subsequently maintained in the laboratory by serial passage of blood from donor infected mice to naive one via intra-peritoneal (IP) route.

2.6 Determination of Body Weight of the Mice

The body weights of each mouse in all groups were measured before and after infections and treatments using sensitive digital weighing balance [13].

2.7 Innoculation of Mice

The parasitaemia of the donor mice were ascertained to be within the range of 20-30%. Blood were collected retro-orbitally and diluted in phosphate buffered saline based on their parasitaema level such that 1mL contains 5 x 10^7

parasites. Each experimental mouse was given 0.2mL of diluted infected blood containing 1 x 10^7 parasitized red blood cells (pRBCs) intraperitoneally.

2.8 Grouping of Experimental Animals

55 Albino mice weighing 20–30g of about 6–8 weeks old purchased were randomly grouped into 11 groups;

Group A – Positive Control (infected and treated with standard drugs)

Group B – Negative Control (infected and not treated)

Group C – 200mg/kg of Azadirachta indica Group D – 400mg/kg of Azadirachta indica Group E – 600mg/kg of Azadirachta indica Group F – 200mg/kg of Senna occidentalis Group G – 400mg/kg of Senna occidentalis Group H – 600mg/kg of Senna occidentalis Group I – 200mg/kg of Momordica balsamina. Group J – 400mg/kg of Momordica balsamina. Group K – 600mg/kg of Momordica balsamina.

After inoculation, each mouse was administered 0.2ml each of the drug (Group A) or extract (Group C – K) orally with canulla according to the groupings for 4 days starting from Day 3 (D3) post infection except the negative control which was given distilled water only. Survival was monitored daily till the completion of the experiment at day 29.

2.9 Preparation of the Extracts and Drugs

Stock solution of each plant extracts was prepared by dissolving in distilled water and stored in an air-tight container in the refrigerator at 4°C. 200mg/kg, 400mg/kg and 600mg/kg were consequently prepared from the stock solution by serial dilution and administered to experimental animal. Piperaquine and dihydroartemisin was dissolved to 32mg and 4mg respectively.

2.9.1 Collection of blood samples for determination of parasitaemia and chemosuppression

Thin blood smears were made from blood collected from the tail of each mouse. The cutting was considerable at small inches so as not to cause prolonged damage to each mouse tail. The slide was fixed in methanol, giemsa stained and viewed under the light microscope. The total number of erythrocytes and total number of infected erythrocytes was calculated in random 5 fields from each slides. Percent parasitaemia and chemosuppression were calculated by:

Percentage parasitaemia = (Number of infected erythrocytes/Total number of erythrocytes) X 100

Percentage chemosuppression = $\frac{(A - B)}{A} \times 100$

Where

A – parasitaemia of negative group

B – parasitaemia of each of treatment/positive group.

2.10 Statistical Analysis

Parasitaemia were analyzed using One-way Analysis of Variance. The statistical significance was defined as P-value of <0.05 using SPSS version 16.0. The differences between the groups were tested using Duncan Post-hoc test. Graph Pad Prism software was used for preparation of Kaplan-Meier survival curves and parasitemia curves.

3. RESULTS

3.1 Yield of Extract

The ethanol extract of all the plants yielded green solid gummy with a percentage yield of 6.5%, 7.6% and 5.7% accounting for 260g, 210g and 350g was obtained from the dried pounded leaves of *Azadirachta indica*, *Momordica balsamina* and *Senna occidentalis* respectively. Result is presented in Table 1.

3.1.1 Phytochemical components detected in plant extracts

The qualitative phytochemical analysis of the ethanolic plant extracts (Table 2) showed the presence of alkaloids, flavanoids, saponins, steroids, phenolics, tannins in all plants. Reducing sugar was present in *Azadirachta indica* and *Senna occidentalis* while terpenoids was present in *Azadirachta indica* and *Momordica balsamina*.

3.1.2 Results of the antiplasmodial activity of ethanolic extract of individual leaves of *A. indica, S. occidentalis* and *M. balsamina* on mice infected with *Plasmodium berghei*

The antiplasmodial activity of the individual plants in the various concentrations showed a

reduced parasitaemia level during the early phase of infection in contrast to the rapid increase observed in negative group. The extracts group produced a parasitaemia count reduction across the day.

On day 6, A. indica showed highest mean parasitaemia of 6.2% in 200mg/kg extract group and 400mg/kg and 600mg/kg extract group showed similar mean parasitaemia of 3.3% (Fig. 1). S. occidentalis had the highest mean parasitaemia of 9.5% in the 200mg/kg extract group, least mean parasitaemia of 4.2% was observed in the 600mg/kg extract group and 400mg/kg extract group had 4.7 % (Fig. 2), M. balsamina had the highest mean parasitaemia of 4.4 % in 200mg/kg extract group, least mean parasitaemia of 3.2 % in the 400mg/kg extract group and mean parasitaemia of 4.3 % in the 600mg/kg extract group (Fig. 3). On the same dav. the negative group had a mean parasitaemia of 7.0 % while the positive group showed a mean parasitaemia of 4.2%.

On day 10, the mean parasitaemia of the negative group had increase to 15.50% while the positive group had a mean parasitaemia of 1.30%. The extracts group showed improved parasitaemia levels with *A. indica* highest mean parasitaemia of 7.5% in 200mg/kg extract group, least mean parasitaemia of 6.2% in 400mg/kg

extract group and a mean parasitaemia of 7.2 % in 600mg/kg extract group, *S. occidentalis* had the highest mean parasitaemia of 5.20% in 200mg/kg extract group, least mean parasitaemia of 3.10% in 600mg/kg extract group and a mean parasitaemia of 4.9 % in 400mg/kg extract group, *M. balsamina* had the highest mean parasitaemia of 5.70% in both 200mg/kg and 600mg/kg extracts group and the least mean parasitaemia of 5.40% in 400mg/kg extract group.

On day 10, the percentage chemosuppression of the positive group was 91.6 % while the negative group had 0.0%. A. indica had the highest percentage chemosuppression of 60.0 % in the 400mg/kg extract group, least percentage chemosuppression of 51.6 % in the 200mg/kg extract group and percentage chemosuppression of 54.0 in the 600mg/kg extract group. S. occidentalis had the highest chemosuppression of 80.0% in the 600mg/kg extract group, least percentage chemosuppression of 66.5% in the 200mg/kg extract group and a percentage chemosuppression of 68.4% in the 400mg/kg extract group. M. balsamina had the highest chemosuppression of 65.1% in the 400mg/kg extract percentage group and а chemosuppression of 63.2 % in both 200 mg/kg and 600mg/kg extracts group (Table 3).

Table 1. Yield of plant extracts of Azadirachta indica, Senna occidentalis and Momordica			
balsamina			

Plants	Quantity of plant materials	Yield of extracts	Percentage yield (%)
A. indica	260g	17g	6.5
M. balsamina	210g	16g	7.6
S. occidentalis	350g	20g	5.7

 Table 2. Phytochemical components identified from crude extract of Azadirachta indica, Senna occidentalis and Momordica balsamina

Phytochemical	Plant extracts		
components	Azadirachta indica	Senna occidentalis	Momordica balsamina
Alkaloids	+	+	+
Flavanoids	+	+	+
Reducing Sugar	+	+	-
Saponins	+	+	+
Steroids	+	+	+
Phenolics	+	+	+
Tannins	+	+	+
Terpenoids	+	-	+

Keys +: Present -: Absent

Bature et al.; JAMB, 21(9): 39-49, 2021; Article no.JAMB.73467

The survival proportion of mice treated with individual plant extracts showed a 100% survival rate at day 6. On day 10, the survival proportions of the extracts group had changed with A. indica highest survival proportion of 100% in 600mg/kg extract group, least survival proportion of 81.8% in 200mg/kg extract group and a survival proportion of 94.7% in 400mg/kg extract group (Fig 4). S. occidentalis had the highest survival proportion of 100.0% in 600mg/kg extract group, least survival proportion of 90.0% in 400mg/kg extract group and a survival proportion of 95.0% in 200mg/kg extract group (Fig 5). Lastly, M. balsamina had the highest survival proportion of 100% in 600mg/kg extract group and the least survival proportion of 89.4% in 400mg/kg extract group. The 200mg/kg extract group had a survival proportion of 95.0% (Fig 6). In comparison, the negative group had the survival proportion of 71.4 % which is significantly lower than the positive group which had survival proportion of 95.0% and considerably lower than the extract groups as discussed above.

3.1.3 Effect of extract on Body weight and rectal temperature

The body weight of the mice in all group were measure on Day zero (0) and Day five (4) before and after treatment respectively. The result of the body weight and rectal temperature are represented in Table 4 and Table 5 respectively.

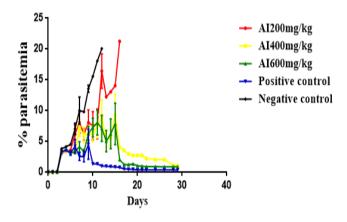


Fig. 1. Effect of the ethanolic leaf extract of *A. indica* on parasitaemia of *P. berghei* infected mice

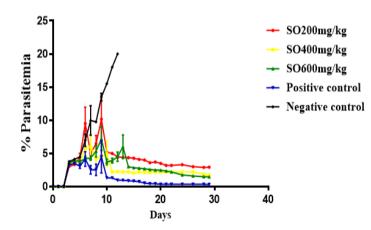


Fig. 2. Effect of ethanolic leaf extract of *S. occidentalis* on parasitaemia of *P. berghei* infected mice

Bature et al.; JAMB, 21(9): 39-49, 2021; Article no.JAMB.73467

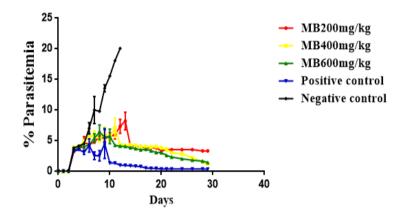


Fig. 3. Effect of ethanolic leaf extract of *M. balsamina* on parasitaemia of *P. berghei* infected mice

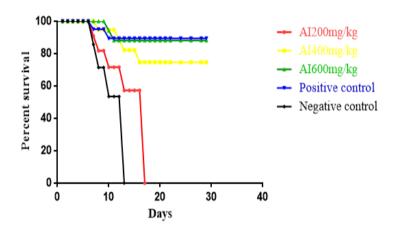


Fig. 4.Survival proportion of *P. berghei* infected mice treated with ethanolic leaf extract of *A. indica*

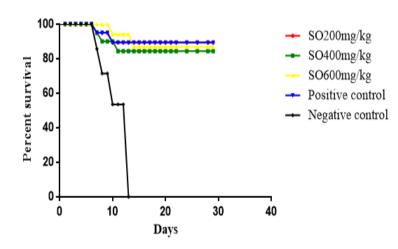


Fig. 5. Survival proportion of *P. berghei* infected mice treated with ethanolic leaf extract of *S. occidentalis*

Bature et al.; JAMB, 21(9): 39-49, 2021; Article no.JAMB.73467

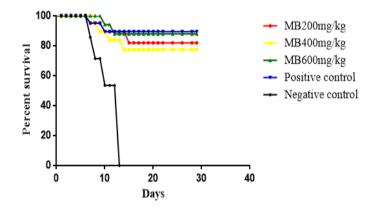


Fig. 6. Survival proportion of *P. berghei* infected mice treated with ethanolic leaf extract of *M. balsamina*

Table 3. Antiplasmodial activity of ethanolic leaf extract of individual plants in mice infected with *P. berghei* on Day 10

Extracts	Dose	% Chemosuppression
Positive	Standard Drugs	91.6
Negative	Distilled water	0.00
A. indica extract	200mg/kg	51.6
	400mg/kg	60.0
	600mg/kg	54.0
S. occidentalis extract	200mg/kg	66.5
	400mg/kg	68.4
	600mg/kg	80.0
M. balsamina extract	200mg/kg	63.2
	400mg/kg	65.1
	600mg/kg	63.2

Table 4. Body weight of mice before and after infection and treatment

Doses	Before	After
Group A – Positive Control (infected and treated with standard	24.00 ± 1.34	22.70 ± 1.33
drugs)		
Group B – Negative Control (infected and not treated)	24.48 ± 0.71	22.90 ± 0.56
Group C – 200mg/kg of Azadirachta indica	26.08 ± 1.11	24.10 ± 1.26
Group D – 400mg/kg of Azadirachta indica	26.12 ± 1.16	24.54 ± 1.10
Group E – 600mg/kg of Azadirachta indica	25.66 ± 1.03	24.24 ± 1.05
Group F – 200mg/kg of Senna occidentalis	27.04 ± 1.41	25.46 ± 1.38
Group G – 400mg/kg of Senna occidentalis	25.50 ± 1.64	24.22 ± 1.71
Group H – 600mg/kg of Senna occidentalis	25.54 ± 0.77	23.88 ± 0.73
Group I – 200mg/kg of Momordica balsamina.	22.90 ± 0.74	21.78 ± 0.67
Group J – 400mg/kg of Momordica balsamina.	25.42 ± 1.33	23.70 ± 1.24
Group K – 600mg/kg of Momordica balsamina.	25.28 ± 1.62	24.10 ± 1.47

Data is presented as Means \pm S.E (n = 5)

4. DISCUSSION

In this study, the yield of *M. balsamina* was high compared to the yield of *S. occidentalis* and *A. indica.* This could be attributed to the pigment-solvent interaction of *M. balsamina* as the plant is known as an important green leafy vegetable

which is a source of important nutrients and supplements according to [14].

This finding is similar to that of [15] whose yield was also high at 11.7% in *Momordica charantia L*. when compared to the extraction of [16] whose *S. occidentalis* yield was low at 7.7%.

Doses	Before	After
Group A – Positive Control (infected and treated with standard drugs)	35.26 ± 0.29	35.26 ± 0.29
Group B – Negative Control (infected and not treated)	37.34 ± 0.26	33.80 ± 0.24
Group C – 200mg/kg of Azadirachta indica	37.62 ± 0.28	34.26 ± 0.22
Group D – 400mg/kg of Azadirachta indica	36.76 ± 0.17	34.30 ± 0.16
Group E – 600mg/kg of Azadirachta indica	37.28 ± 0.11	36.04 ± 0.34
Group F – 200mg/kg of Senna occidentalis	36.98 ± 0.28	33.42 ± 0.17
Group G – 400mg/kg of Senna occidentalis	37.40 ± 0.31	34.62 ± 0.17
Group H – 600mg/kg of Senna occidentalis	37.76 ± 0.19	36.78 ± 0.13
Group I – 200mg/kg of <i>Momordica balsamina</i> .	37.46 ± 0.14	34.22 ± 0.15
Group J – 400mg/kg of Momordica balsamina.	38.08 ± 0.04	36.36 ± 0.17
Group K – 600mg/kg of Momordica balsamina.	37.12 ± 0.34	35.62 ± 0.42

Table 5. Rectal temperature of mice before and after infection and treatment

The quantitative phytochemical screenings of the three plants indicate the presence of alkaloids. flavanoids, saponins, steroids, phenols, and tannin which have been suggested to confer antiplasmodial activity [4]. This is similar to the findings of [17] who confirmed the presence of tannins, flavonoids, alkaloids, saponins, polyphenols and steroids in A. indica, also the findings of [18] confirmed the presence of alkaloids, flavonoids, phenols, steroids. saponins, tannins and anthroquinones in Cassia occidentalis and the findings of [19] also confirmed the presence of tannins, flavonoids, alkaloids and saponins in M. balsamina.

Alkaloids are among the major natural products known to possess antimalarial and various classes of it have been reported to exhibit reliable actions. [20]. The presence of this compound therefore may be responsible for the antiplasmodial activity observed in the plant extract. [21] significantly describe quinine to be a synthetic derivative of the alkaloids. The presence of secondary metabolites like saponins and flavonoids have also been reported to confer antimalarial activity. [22, 23].

The individual study of the plant extract showed that the treatment groups were significant different (P < 0.05) from that of the negative control group at day 6 in terms of parasitaemia growth inhibition. Even though the levels of the parasitaemia increase along the days, these were still lower than that observed in the negative group demonstrating the presence of inhibitory properties in them.

The extracts plant being antioxidant may have resulted in free radical scavenging and lipid peroxidation inhibition [24], thus conferring adequate protection enough to counter the effect of the oxidative stress induced in host parasitized erythrocyte by the malaria parasite [25]. Typically, malaria parasite digest haemoglobin in vacuole to amino acids and heme [26], therefore one of the most important mechanism in its prevention is inhibition of heme polymerization. This inhibition is exhibited by the antioxidant activity before polymerization, and the unpolymerized heme is very toxic for the intraerythocytic plasmodial[27].

The individual plant extract with the highest effect is the *S. occidentalis* 600mg/kg stating adequacy in its ability to confer antiplasmodial ability. This may be attributed with the presences of phytochemical constituents revealed in the phytochemical studies of the plant [28].

Reduction in body weight, anaemia and rectal temperature decrease are typical features of malaria in mice. [29]. This is closely concerned with the clearance of infected and uninfected RBCs and erythopoises inhibition by malaria parasite. [30].

The *S. occidentalis* although confer 80% chemosuppresion while the standard drugs had 91.6% chemosuppression. This substantiates the necessity to determine other details like pharmacokinetic and pharmacodynamics of the plant extracts.

5. CONCLUSION

In conclusion, the extraction solvent yields enough extract of the three plants for the research. The preliminary screening revealed the presence of important phytochemical like alkaloid, flavonoid, phenols, steroids, saponins and tannin in the plants. The individual study of plants extracts had lower percentage parasitaemia and therefore conferred more antiplasmodial activity. This *invivo* study of the plant extracts therefore demonstrated activity which have individually resulted in prolonged survival of the mice in the groups.

ETHICAL APPROVAL

The experiment management, Animal handling and care were approved by the Research and Ethics Committee of the Department of Biological Sciences, Nigeria Defence Academy, Kaduna, Kaduna State.

ACKNOWLEDGEMENT

Profound appreciation goes to my parents who have contributed morally and financially to the success of this research. My sincere appreciation goes to my supervisor, my H.O.D., lecturers and laboratory assistant for their understanding, words of encouragement, intense research corrections and guides. The Project was funded completely by Author AMB.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. White NJ. Malaria time to act. The New England Journal of Medicine. 2006;355:1956 1957.
- 2. Wang F, Feng Y, Pang W, Qi Z, Zhang Y, Guo Y, Luo E, Cao Y. Parasite densities modulate susceptibility of mice to cerebral malaria during co- infection with *Schistosoma japanicum* and *Plasmodium berghei*. Malaria Journal. 2014;13: 116.
- Desai M, Ter Kuile FO, McGready R, Asamoa K. Epidemiology and burden of malaria in pregnancy. Lancet Infectious Disease. 2007;7:93-104.
- 4. Tarkang PA, Okalebo FA, Ayong LS, Agbor GA, Guantai AN. Anti-malarial activity of a polyherbal product (Nefang) during early and established *Plamodium* infection in rodent models. Malaria Journal. 2014;13:456.
- Bandaranayake WM, Ahmad I, Aqil F, Owais M. 'Quality control, screening, toxicity, and regulation of herbal drugs', *Modern Phytomedicine*. Turning Medicinal Plants into Drugs. 2006;25 – 57.
- 6. Ogbuewu IP, Odoemenam VU, Obikaonu HO, Opara MN, Emanalom OO, Uchegbu

MC, Okoli IC, Esonu BO, Iloeje MU. the growing importance of Neem (*Azadirachta indica* A. Juss) in Agriculture, Industry, Medicine and Environment. Research Journal of Medicine Plants. 2011;5 (3):230 – 245.

- Bijauliya RK, Alok S, Chanchal DK, Sabharwal M, Singh M. An updated review of pharmacological studies on *Azadirachta indica* (neem). International Journal Pharmaceutical Sciences and Research. 2018;9(7):2645 – 55.
- Ibrahim MA, Aliyu AB, Sallau AB, Bashir I, Yunusa I, Umar TS. Senna occidentalis leaf extract possesses antitrypanaosomal activity and ameliorated the trypanosomeinduced anemia and organ damage. Pharmacognosy Reseach. 2011;2(3):175 – 180.
- Kaur I, Ahmad S, Harikumar SL. Pharmacognosy, Phytochemistry and Pharmacolgy of *Cassia occidentalis* Linn. International Journal of Pharmacognosy and Phytochemical Research. 2014;6(2):151 – 155.
- Thakur GS, Bag M, Sanodiya BS, Bhadauriya P, Debnath M, Prasad GBKS, Bisen PS. *Mormodica balsamina*: A Medical and Neutraceutical Plant for Health Care Management. Current Pharmaceutical Biotechnology. 2009;10(7):667 – 682.
- Anokwuru C. Effect of Extraction Solvents on Phenolic, Flavonoid and Antioxidant activities of three Nigerian medicinal Plants. Nature and Science. 2011;9(7):53 – 61.
- Das BK, Al-Amin MM, Russel SM, Kabir S, Bhattacherjee R, Hannan JMA. Phytochemical Screening and Evaluation of Analgesic Activity of *Oroxylum indicum*. Indian Journal Pharmaceutical Sciences. 2014;76(6):571–575.
- Dada EO, Oloruntola DA. In vivo antiplasmodial activity of ethanolic leaf extract of *Tithonia diversifolia* (Hemls.) A. Gray against *P. berghei* NK65 in infected Swiss Albino Mice. Journal of Applied Life Science Internationsl. 2016;8(3):1 – 8.
- Hassan LG, Umar KJ. Nutritional Value of Balsam Apple (*Momordica balsamina* L.) leaves. Pakistan Journal of Nutrition. 2006;5(6):522 – 529.
- Baonda SM, Loumpangou CN, Ampa R, Ndinga AME, Milandou LJ, Bonose CM, Ouamba JM. Evaluation of antidiabetic activity of the ethanol extract of *Momordica*

charantia, L. and the identification of charantine by gas chromatography coupled with Mass spectrometry. Journal of Medicinal Plants Research. 2019;13(14):321 – 328.

- Adamu U, Abdulrazak MH, Yusha'u M, Salisu B. Antibacterial potentials and toxicity study of *Cassia occidentalis* leaf extracts against clinical isolates of *Salmonella sp.* Science World Journal. 2018;13(1):77 – 81.
- Ravi K, Bharavi K, Ravi Kumar P, Krishna B. Phytochemical and Antibacterial activity of ethanol extract of leaves and twigs of *Azadirachta indica A. Juss.* United Kingdom Journal of Pharmaceutical and Biosciences. 2015;3(6):56 – 59.
- Srividya S, Sridevi G, Manimegalai AG. Phytochemical Screening and *In Vitro* antioxidant activity of ethanolic extract of *Cassia occidentalis*. International Journal of Pharmaceuticals and Clinical Research. 2017;9(3):252–256.
- Abubakar B, Fatima M, Sulaiman SK, Bashir A, Buhari Y. Antimicrobial activity of Balsam Apple (*Momordica balsamina L.*). UMYU Journal of Microbiology Research. 2018;3(1):24 – 29.
- 20. Oliveira AB, Dolabela MF, Braga FC, Jacome RL, Varotti FP, Povoa MM. Plantderived antimalarial agents: new leads and efficient phythomedicines. Part I Alkaloids. Academia Brasileira de Ciencias. 2009;81:715–740
- 21. Cassani C, Martin-Rapun R, Arceo E, Bravo F, Melchiorre P. Synthesis of 9amino (9-deoxy) *epi* cinchona alkaloids, general chiral organocatalysts for the stereoselective functionalization of carbonyl compounds. Nature. 2013;8:325-344.
- 22. Oladeji OS, Odelade KA, Oloke JK. Phytochemical screening and antimalarial investigation of *Moringa oleifera* leaf extracts. African Journal of Science,

Technology, Innovation and Development. 2020;12(1):79 – 84.

- 23. Gupta M, Mazumder P, Gomathi P, Selva VT. Antimalarial activity of methanol extract of *Plumeria acuminate* Ait, leaves and *Tephrosia purpurea* (Linn) Pers Root. Natural Radiance. 2008;7(2):103 –105.
- 24. Orabi KY, Al-Qasoumi SI, El-Olemy MM, Mossa JS, Muhammad I. Dihydroagarofuran alkaloid and triterpenes from Maytenus heterophylla and Maytenus arbutifolia. *Phytochemistry*, 2001;58: 475-480.
- Percario S, Moreira DR, Gomes BAQ, Ferreira MES, Goncalves ACM, Laurindo PSO, Vilhena TC, Dolabela MF, Green MD. Oxidative stress in malaria. International Journal of Molecular Sciences. 2012;13(12):16346-16372.
- Esmaeili, S., Irani, M., Moazzeni, H. and Mosaddegh, M. (2020): Inhibition of heme polymerization, the mechanism of antimalarial activity in *Phlomis caucasica* Rech. F. (Lamiacaea). *Journal of Medicinal Plants.* 18(72): 103 – 109.
- 27. Somsak V. *In Vivo* Antimalarial activity of *Annona muricata* Leaf Extract in Mice Infected with *Plasmodium berghei*. Journal of Pathogens. 2016;3264070:1 – 5.
- Saidu AN, Aina EO, Mann A, Leje UI. the effect of aqueous extract of Senna occidentalis leaves on rats infected with Salmonella typhi. Austraillian Journal of Basic and Applied Sciences. 2011;5(12):1863 -1867.
- 29. Langhorne L, Stuart QJ, Sanni LA. Mouse Models of blood-stage malaria infections: Immune responses and cytokines involved in protection and patology. Malaria immunology. 2002;80 (80):204 – 228.
- 30. Wubetu YB, Abyot EG, Zewdu BW.Antimalaria activity of stem bark of *Periploca linearifolia* during early an established *Plasmodium* infection in mice;2018. ID 4269397.

© 2021 Bature et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle4.com/review-history/73467